

**Trends in Phytochemical Research (TPR)** 



Journal Homepage: http://tpr.iau-shahrood.ac.ir

Review Paper

# A review of <sup>13</sup>C NMR spectra of drimane sesquiterpenes

# SYLVIA AWINO OPIYO

<sup>1</sup>Department of Physical and Biological Sciences, Murang'a University of Technology, P. O. Box 75-10200, Murang'a, Kenya

# ABSTRACT

Drimane sesquiterpenes are important group of phytochemicals with a wide range of medicinal properties including antibacterial, antifungal, anti-inflammatory, antioxidant, antiplasmodial, antileishmanial, molluscicidal, antifeedant and insecticidal features. Due to their wide range of medicinal activities, scientists are prompted to continuous-ly search for novel drimane sesquiterpenes since most of the currently available anti-inffective agents have drawbacks such as drug resistance and side effects. Structure determination of new compounds relies on accurate interpretation of spectroscopic data which is quite challenging but can be simplified by comparison with the data of known related compounds from the reliable databases. A large number of drimane sesquiterpenes have been examined by <sup>13</sup>C NMR spectroscopy but such data are scattered in the literature making it hard for researchers to use them. This paper provides a review on previously reported drimane sesquiterpenes and a compilation of their <sup>13</sup>C NMR data. It also provides a brief discussion on the substituent effect on the <sup>13</sup>C shielding of the drimane sesquiterpenes.

© 2019 Islamic Azad University, Shahrood Branch Press, All rights reserved.

# 1. Introduction

The use of medicinal plants as health remedies has been practiced for several thousands of years. In many developing countries, traditional medicine still plays an important role in meeting the primary healthcare needs of the population (WHO, 2014; Mohammadhosseini et al., 2019). Plants produce secondary metabolites that have the capacity to combat numerous types of diseases (Jeruto et al., 2017; Mohammadhosseini et al., 2017; Mohammadhosseini, 2017). Despite the availability of conventional drugs, continued search for novel biologically active compounds is unavoidable since most of the available drugs have demonstrated limitations in terms of side effects and drug resistance (WHO 2014). Traditional remedies are preferred because they are cheaper and readily available (WHO 2014). In recent years, many researchers have focused on authenticating the efficacy of medicinal plant extracts through in-vivo and in-vitro experiments, and isolation and structural determination of the bioactive compounds (Opiyo et al., 2017; Wansi et al., 2018; 2019). This has led to the identification of several important biologically active compounds including terpenes, alkaloids, steroids, flovonoids and quinones (Ochieng et al., 2013; Ochieng et al., 2017; Ochung et al., 2015; 2018). Such biologically active compounds derived from natural origin represent an important source of drugs in the process of developing new pharmacologically active compounds (Vieira et al., 2014). Drimane sesquiterpenes (1a) are C-15 compounds that occur as hydrocarbons or in oxygenated forms such as alcohols, ketones, aldehydes, acids or lactones in nature. Rearranged (1b), tricyclic (1c) as well as 11-*nor* (1d) and 12-*nor* (1e) drimanes sesquiterpenes also occur in

nature (Fig. 1). Drimane sesquiterpenes are important constituents of essential oils which have various physiological effects. Most of these compounds have nice aroma and are used in soap and perfumery industry as

# ARTICLE HISTORY

Received: 01 May 2019 Revised: 21 June 2019 Accepted: 22 August 2019 ePublished: 22 September 2019

#### K E Y W O R D S

Chemical shift <sup>13</sup>C NMR data Drimane sesquiterpenes Shielding Structure determination



well as flavor compounds in aroma mixtures.



Fig. 1. Structures of drimane sesquiterpenes.

Drimane sesquiterpenes have attracted particular interest because of their numerous potent medicinal properties which include antibacterial (Wube et al., 2005; Opiyo et al., 2011), antifungal (Delgado et al., 2008), antimycobacterial (Wube et al., 2005; Madikane et al., 2007), anti-inflammatory (Cunha et al., 2001; Felix et al., 2014), antioxidant (Jansen and de Groot, 2004), antiplasmodial (Wube et al., 2010; Pittayakhajonwut et al., 2011; Claudino et al., 2013; Nyaba et al., 2018), antileishmanial (Claudino et al., 2013), antifeedant (Kubo and Nakanishi, 1977; Kubo, 1995; Chaudhary et al., 2008; Montenegro et al., 2013; 2018a 2018b; Inocente et al., 2018), molluscicidal (Nakanishi and Kubo, 1978; Kubo et al., 1983; Odyek et al., 1993; Montenegro et al., 2014) and insecticidal properties (Muñoz et al., 2015; Opiyo et al., 2015; Inocente et al., 2018; Montenegro et al., 2018).

1.1. Occurrence of drimane sesquiterpenes

# Table 1

Some plants sources of drimane sesquiterpenes.

Drimane sesquiterpenoids show a restricted occurrence in a few unrelated lower and higher plants including Canella winterana (L.), Cinnamosma fragrans (Baill) and Warburgia ugandensis Sprague (Table 1). The compounds have been isolated from fungi such as Phellinus tuberculosis (Quél.), Marasmius oreades (Bolton) Fr., Aspergillus sp. (Micheli.), Kuehneromyces sp. (Singer & Å.H. Sm.), Trichopezizella barbata (Kunze Fr.), Mniopetalum sp. (Pers.) Penicillium brevicompactum Dierckx, Lactarius uvidus (Fr.), Polyporus ciliates (Fries), P. arcularius Rostk. Pestalotiopsis spp. (Steyaert) and Lepista glaucocana (Bres.) (Xu et al., 2009a; Felix et al., 2014; He et al., 2015; Chen et al., 2016; Ding et al., 2016; Kunag et al., 2016; Zhao et al., 2017). Drimane sesquiterpenes have also been isolated from marine sponges (Butler and Capon, 1993; Montagnac et al., 1996; Paul et al., 1997). Synthetic drimane sesquiterpenoids have been reported by various coworkers (Lagnel et al., 2000; Jansen and de Groot, 2004; Vlad et al., 2006), as well.

#### 1.2. Structure elucidation

Structure determination of new natural products relies majorly on the acquisition and accurate interpretation of spectroscopic data (Mahato and Kundu, 1994). The assignment of carbon signals of a new compound by comparison with the data of known compounds is sim-

Family Amaranthaceae	Plant species Tidestromia oblongifolis (S. Watson) Standl	Reference Chaudhary et al., 2008
Apiaceae	Heptaptera anatolica (Boiss.) Tutin Heptaptera anisoptera Tutin	Tosun et al., 2019 Appendino et al., 1992
Asteraceae	Tanacetum heterotomum (Bornm.) Tanacetum parthenium L.	Gören and Ulubelen, 1988 Kisiel and Stojakowska, 2002
Canellaceae	Canella winterana (L.) Gaertn. Cinnamodendron corticosum (Miers) Cinnamosma fragrans (Baill) Warburgia salutaris (Bertol. f.) Chiov Warburgia stuhlmannii Engl. Warburgia ugandensis Sprague Capsicodendron dinisii (Schwacke) Occhioni	Grace et al., 2010 Seeram et al., 2003 Inocente et al., 2018 Nyaba et al., 2018 Kioy et al., 1990 Opiyo et al., 2011 Bastos et al., 1999
Cinnamosma Makinoaceae Polygonaceae	Cinnamosma madagascariensis Danguy Makinoa crispate (Stephani) Miyake Polygonum hydropiper Linn. Polygonum glabrum Willdenow Polygonum punctatum (Elliott)	Harinantenaina et al., 2008 Hashimoto et al., 1989 Sultana et al., 2011 Jacobsson and Muddathir, 1992 Alves et al., 2001
Porellaceae	Porella acutifolia (Lehm. & Lindenb.) Trevis Porella canariensis (F. Weber) Underw Porella cordeana (Hiib.) Evs. Porella navicularis (Lehm. & Lindenb.) Lindb. Porella vernicosa Lindb.	Ludwiczuka et al., 2011 Ludwiczuka et al., 2011 Ludwiczuka et al., 2011 Ludwiczuka et al., 2011 Ludwiczuka et al., 2011
Scapaniaceae Solanaceae	Diplophyllum serrulatum (K. Müller) Nicotiana tabacum L.	Toyota et al., 1994 Hlubucel et al., 1974
Umbelliferae	Ferula assa-foetida L. Ferula gummosa Boiss.	Lee et al., 2009 Iranshahi et al., 2014
Winteraceae Zingiberaceae	Drimys arfakensis (Gibbs.) Drimys brasiliensis Miers Drymus winteri (J.R. Forst. & G. Forst.) Pseudowintera colorata (Raoul) Dandy Pseudowintera insperata Heenan et de Lange Tasmannia lanceolata R.Br. Hedychium spicatum (Buch.) Ham.	Santoso et al., 2018 Claudino et al., 2013 Muñoz et al., 2015 Wayman et al., 2010 Wayman et al., 2010 Mathie et al., 2017 Reddy et al., 2009



10 7a, 8a, 11-OH

11 6a, 8a, 11-OH

12 9a, 11, 12-OH

13 8a, 11, 12-OH

15 3β, 8β, 11, 12 -OH, sulphureuine B

16 3α, 8β, 11, 12-OH, sulphureuine C

17 3 $\beta$ , 11, 12-OH, sulphureuine G

11, 12-OH

ple and straightforward provided that the <sup>13</sup>C NMR data of appropriate model compounds are available. A large number of drimane sesquiterpenes have been examined by <sup>13</sup>C NMR spectroscopy and considerable <sup>13</sup>C NMR chemical shift data have been accumulated. However, these data are scattered in the literature making it hard for researchers to use them. It is therefore necessary to provide an easy access to an extensive list of <sup>13</sup>C data of drimane sesquiterpenes.

# 2. <sup>13</sup>C NMR data of drimane sesquiterpenes

This paper provides a compilation of <sup>13</sup>C NMR data of selected varieties of naturally occurring and synthetic drimane sesquiterpenes that were previously reported. The sesquiterpenes have been arranged skeleton-wise in the following order: saturated drimane sesquiterpenes **2-24**; drim-8(12)-ene sesquiterpenoids **25-34**; drim-7-ene sesquiterpenes **35-80**; drim-8 (9)-ene sesquiterpenes **81-93**; epoxydrimane derivatives **94-101**; drim-6,8-diene derivatives **102-103**; drim-5,8-diene **104-105**; rearranged drimane sesquiterpenes **106-109**; tricyclic drim-7-enes **110-126**, tricyclic drim-8(9)-enes **127-143**; tricyclic drim-2,8-dienes **144-149**; miscellaneous unsaturated tricyclic drimane sesquiterpenes **150-152**;

#### Table 2

Structures of drimanes 2-24.



For cpds 2, 3, 5-12, 14-24

2 8α-OH 3 8β-OH, driman-8β-ol 4 8β-OH, isodriman-8β-ol 5 7β, 8β, 9α-OH 6 11-OH, 14 3β, 8α, 7 11, 12-OH 8 8α, 11-OH 9 7β, 8β, 11- OH

Non-substituted drimane sesquiterpene (1a) consists of five methyl (C-11, C-12, C-13, C-14 & C-15), five methylene (C-1, C-2, C3, C-6 & C-7), three methine (C-5, C-8 & C-9) and two quaternary (C-4 & C-10) carbon atoms. The <sup>13</sup>C NMR chemical shifts for all the 15 carbon in a non-substituted drimane range between  $\delta$  7-60 ppm. Substitution affects the chemical shifts of adjacent and nearby nuclei. Inspection of the <sup>13</sup>C NMR data of various mono- and polyhydroxy drimanes sesquiterpenes (Table 3) reveals that introduction of a hydroxyl group results in downfield shifts of 34-50 ppm for  $\alpha$ -carbons and 2-10 ppm for  $\beta$ -carbons and upfield shifts of 0-9 ppm for  $\gamma$ -carbons. Generally, hydroxylated carbon atoms at

saturated tricyclic drimane sesquiterpenes **153-158**; rearranged tricyclic drimane sesquiterpenes **159-162** and nordrimane derivatives **163-183**.

### 2.1. Saturated drimane sesquiterpenes

Several groups of researchers have assigned the <sup>13</sup>C resonances of a variety of saturated drimane sesquiterpene consisting of monohydro 2-4, 6, 18, 21, dihydro 7, 8, trihydro 5, 9-13, 15-17, and tetrahydro drimane derivatives (Table 2). The <sup>13</sup>C NMR values of representative saturated drimanes 2-24 are summarized in Table 3 (Montagnac et al., 1996; Meng et al., 2011; Yonemura et al., 2012; Derita et al., 2013; Skiredj, 2016). Most of the compounds have hydroxyl groups attached to C-6, C-7, C-8, C-11 and C-12 (Barrero et al., 1995; Lagnel et al., 2000; Benites et al., 2001; Panasenko et al., 2004; Yonemura et al., 2012; He et al., 2015; Skiredj, 2016). Compounds 14-17 have hydroxyl group attached to C-3 (Meng et al., 2011; He et al., 2015). Mono and di-acetylated derivatives 19, 21-24 have been reported. In most cases, the acetyl substitution occurs at C-8, C-11 and C-12 (Barrero et al., 1995; Lagnel et al., 2000; Benites et al., 2001).



For cpds 4 & 13

18 8α-OH, 11-CHO 19 6α, 8α-OAc, 11-CHO 20 8α-COOH, 11-CHO 21 8α-OH, 11-OAc 22 11, 12-OAc 23 8α-OAc, 11-COOH 24 8α-OAc, 11-COOMe

C-3, C-7 & C-8 resonate between  $\delta$  71-85 ppm, while those at C-6, C-11 & C-12 resonate between  $\delta$  59-70 ppm. In polyhydroxy drimanes such as in 1,2- or 1,3-dihydroxy derivatives, there is additivity of the substituent effect on chemical shifts of the nuclei that explains, for example, why C-8 in compound 5 resonates more downfield than that in isodriman-8 $\beta$ -ol (4). Acetylation of the hydroxyl group enhances the  $\alpha$ -effect and diminishes the  $\beta$ -effect. However, the  $\gamma$ -effect remains more or less unaltered. The effect at  $\beta$ -position is attributed to the  $\gamma$ -effect of the acetyl moiety (Mahato and Kundu, 1994). For example, in driman-8,11-diol (8) and 11-acetoxydriman-8 $\alpha$ -ol (21), C-11, C-9 and C-10 resonate at



 $\delta$  61.0, 60.5, 37.5 and 62.6 (+1.6), 59.4 (-1.1), 38.1 (+0.6) ppm, respectively. The substituent effects depend on the degree of substitution of carbon under consideration as follows: quaternary carbons < methine carbons < methine carbons (Mahato and Kundu, 1994).

The configuration at C-9 in a drimane sesquiterpene can be  $\alpha$  or  $\beta$  and the difference can be observed considering the chemical shift of  $\gamma$ - carbons at C-1, C-5 and C-15 (Rodriguez et al., 2005). For example, driman-8 $\beta$ -

stereogenic center and the configuration of the C-11 methyl substituent is evidenced by the noticeable difference in the chemical shifts of the C-1, C-5 and C-15 $\gamma$  carbons [ $\Delta \delta = \delta(3) - \delta(4)$ : +2.9, +10.5 and -9.6 ppm, respectively]. The presence of 1,3-diaxial interactions results in less shielding of nuclei as in the case of methyl C-11 carbon in compound **4** which results to a downfield shift by ~4.7 ppm (Table 3).

#### Table 3

<sup>13</sup> C NMR data of drimane sesquiterpenes.

C	2c	34	4c	2c	90	7c	8c	9с	10 <sup>c</sup>
1	39.9	40.8	37.9	32.9	39.9	39.4	40.0	40.1	39.6
2	18.7	19.3	18.8	18.3	17.5	18.5	18.6	18.4	18.6
e	42.0	42.8	42.7	41.5	41.9	41.9	41.7	41.7	40.8
4	33.3	33.9	33.2	33.2	33.2	33.2	33.2	33.2	32.7
v	56.2	56.9	46.4	43.7	56.5	56.4	55.9	52.4	45.1
9	20.5	19.3	18.8	27.7	18.4	18.3	20.1	27.7	25.7
7	44.5	43.6	35.6	73.5	34.5	30.0	44.3	76.6	74.0
œ	73.1	71.8	74.9	77.3	28.5	37.9	75.0	75.7	75.3
6	55.6	53.6	53.5	78.9	55.7	54.5	60.5	57.8	54.4
10	37.8	38.7	37.4	42.2	37.6	37.3	37.5	38.1	37.0
11	7.4	7.9	12.6	17.6	61.0	60.4	61.0	59.8	60.8
12	23.1	31.4	32.4	21.8	15.6	66.4	24.2	25.8	23.1
13	33.5	34.0	33.5	21.8	21.6	33.5	33.5	21.6	21.6
14	21.6	22.2	22.0	33.6	33.6	21.6	21.6	33.6	32.3
15	14.4	14.9	24.5	17.1	17.1	16.4	16.0	16.7	15.6
Ref	Wahlberg et al., 1981	Yonemura et al., 2012	Ohloff and Giersch, 1985	Panasenko et al., 2004	Derita et al., 2013	Benites et al., 2001	Barrero et al., 1995	Panasenko et al., 2004	Panasenko et al., 2004



C	11 <sup>c</sup>	12	13 <sup>c</sup>	14 <sup>c</sup>	15 <sup>D</sup>	16 <sup>b</sup>	17 <sup>0</sup>	18 <sup>c</sup>	19 <sup>c</sup>	20 <sup>c</sup>
-	40.2	31.5	37.4	38.1	37.8	32.4	37.5	40.0	43.3	40.2
2	18.4	18.7	18.8	26.6	27.0	25.2	27.1	18.3	17.6	19.0
3	43.1	41.5	42.1	77.8	76.9	73.7	76.7	42.9	39.7	42.5
4	33.7	33.4	33.4	38.6	38.6	37.2	38.6	33.4	33.0	34.0
S	59.8	46.1	56.9	55.3	54.2	49.7	55.0	41.8	56.3	54.9
9	68.6	21.2	20.0	19.2	17.5	17.3	17.2	20.0	69.1	21.8
٢	36.3	26.1	33.1	36.7	36.9	36.9	28.6	40.0	45.6	30.2
×	74.5	42.9	76.4	75.1	73.7	74.0	37.7	71.4	81.9	40.2
6	61.1	75.4	48.6	60.4	54.1	55.9	55.2	55.3	67.4	64.4
10	38.2	41.5	37.6	37.1	37.1	37.1	36.5	37.5	38.2	38.9
11	53.9	63.6	60.9	58.7	57.4	57.4	58.3	208.3	202.0	210.9
12	22.3	64.9	69.5	63.6	68.8	68.8	59.3	25.4	21.0	182.9
13	33.7	22.1	21.8	27.3	16.5	16.5	16.3	33.5	35.5	34.2
14	25.7	33.6	23.7	14.7	16.0	22.2	15.8	21.5	22.9	22.2
15	18.2	15.4	33.0	15.9	28.6	28.9	28.3	17.7	17.6	16.6
16									169.1	
17									21.7	
18									169.1	
19									21.8	
Ref.	Lagnel et al., 2000	Benites et al., 2001	Derita et al., 2013	Meng et al., 2011	He et al., 2015	He et al., 2015	He et al., 2015	Skiredj, 2016	Lagnel et al., 2000	Montagnac et al., 1996

ec	t														
	42.0	33.6	54.8	24.0	37.7	146.7	55.1	39.0	61.6	107.1	33.7	21.9	15.2	171.2	21.2
										0				0	

Opiyo/ Trends in Phytochemical Research 3(3) 2019 147-178

21 <sup>c</sup>	22 <sup>c</sup>	23 <sup>c</sup>	24 <sup>c</sup>	25 <sup>8</sup>	26 <sup>c</sup>	27 <sup>c</sup>	28 <sup>c</sup>	29 <sup>c</sup>	30 <sup>c</sup>
	39.1	39.7	39.7	39.5	42.4	39.1	38.9	37.6	39.1
	18.4	18.2	18.2	19.6	19.3	19.2	19.3	23.6	19.3
	41.8	41.7	41.8	42.4	37.6	41.9	42.0	80.5	42.0
	33.3	33.1	33.1	33.5	33.9	33.5	33.1	38.1	33.6
	56.2	55.1	55.1	55.4	52.6	55.2	47.4	54.5	54.8
	17.6	20.1	20.1	24.2	22.8	24.2	30.4	24.2	24.0
	29.1	38.7	38.9	37.7	36.0	37.9	73.6	36.7	37.7
	34.9	85.0	85.2	151.5	161.5	147.9	148.4	147.1	146.7
	51.4	63.0	63.4	50.5	149.9	59.1	53.2	58.8	55.1
	37.1	38.4	38.6	39.0	40.3	39.0	39.2	38.6	39.0
	62.6	172.5	172.5	10.6	103.0	58.8	58.3	58.8	61.6
	63.9	22.6	22.6	101.5	108.8	106.3	109.9	107.0	107.1
	33.4	33.3	33.2	33.7	22.2	33.6	33.4	28.31	33.7
	21.5	21.3	21.3	22.0	33.5	21.8	21.6	16.7	21.9
	16.4	15.2	15.3	13.5	20.8	15.3	14.4	15.4	15.2
	171.1	170.0	169.6					171.0	171.2
	21.0	20.8	20.9					21.3	21.2
	171.1		50.9						
	21.0								
	3enites et al., 2001	Barrero et al., 1995	Barrero et al., 1995	Yonemura et al., 2012	Kinoshita et al., 2002	Delgado et al., 2008	Barrero et al., 1995	Justicia et al., 2005	Toshima et al., 2001





- C	<b>31</b> <sup>c</sup> 38.6	<b>32</b> <sup>c</sup> 37.2	<b>33</b> <sup>c</sup> 37.3	<b>3</b> 4 <sup>c</sup> 36.5	<b>35</b> <sup>c</sup> 39.4	<b>36<sup>c</sup></b> 36.8	<b>3</b> 7 <sup>c</sup> 34.5	<b>38</b> <sup>M</sup> 38.9	<b>39</b> <sup>c</sup> 39.3	40 <sup>B</sup> 39.8
	0.00	7:10	c./c	C.UC	+.7C	0.00	C:+C	C.0C	C.YC	0.70
	19.2	23.7	23.4	22.7	18.8	21.7	38.5	28.1	18.2	19.3
	42.0	78.8	80.4	80.5	42.2	42.7	216.7	79.6	43.0	42.6
	33.1	38.9	38.1	38.0	32.9	33.0	47.5	39.7	32.3	33.0
	47.2	54.5	54.4	53.9	40.9	57.6	51.1	51.0	58.2	50.4
	30.6	27.9	24.2	24.0	23.6	24.0	23.8	24.2	200.3	24.1
	73.7	27.6	36.7	35.8	123.9	124.6	123.7	123.7	129.0	121.9
	148.2	146.4	146.1	143.1	133.0	131.1	132.9	134.9	157.5	135.3
	49.4	54.7	54.4	62.5	57.3	43.4	56.0	58.2	63.0	49.0
	39.2	39.4	38.6	38.8	36.1	36.1	35.8	36.8	42.1	36.0
	61.4	61.7	61.5	165.6	60.9	61.3	60.6	61.2	59.9	11.6
	110.5	107.8	107.7	108.9	21.9	22.2	21.7	22.2	21.7	22.0
	33.4	28.5	28.30	28.2	33.4	33.2	25.2	28.7	33.6	33.5
	20.4	15.7	16.6	16.6	22.0	23.0	22.3	15.9	22.0	22.1
	14.2	15.5	15.2	14.3	12.8	18.8	14.5	14.9	15.9	13.5
	171.4	171.6	171.2	171.8						
	20.9	21.3	21.3	21.3						
			170.9	51.0						
			21.1							
	Barrero et al., 1995	Dacunto, 2012	Justicia et al., 2005	Justicia et al., 2005	Aasen et al., 1977	Derita et al., 2013	Xu et al., 2009a	Xu et al., 2009a	Lagnel et al., 2000	Yonemura et al., 2012



- c	41 <sup>c</sup>	42 <sup>c</sup> 31.6	<b>43</b> <sup>c</sup> 47 4	44^ 33.7	45 <sup>c</sup> 31.6	46 <sup>c</sup> 37 8	47 <sup>b</sup> 29.6	48 <sup>b</sup>	49 <sup>c</sup> 301	50
-	40.4	31.0	42.4	33.2	31.0	37.8	29.62	41.0	39.1	27.3
7	18.3	18.6	18.4	19.5	18.0	20.4	26.3	62.4	18.1	37.6
9	42.0	41.8	42.6	45.2	42.6	43.1	76.7	51.7	42.9	78.9
4	33.0	32.1	34.6	34.8	32.2	32.8	37.1	33.4	32.2	38.7
S	49.1	42.9	50.3	47.2	55.7	61.1	55.3	54.7	54.6	49.3
9	23.7	24.1	68.4	65.4	200.2	199.6	199.5	199.6	199.6	23.2
٢	125.5	127.5	124.3	129.4	129.2	128.0	128.2	128.1	129.4	123.5
×	127.8	135.3	128.9	137.7	154.9	156.2	157.5	157.6	155.4	132.3
6	67.6	75.6	76.0	75.4	74.7	150.0	74.6	74.6	63.0	53.2
10	37.0	40.7	38.1	41.2	45.0	42.9	44.5	46.2	42.2	35.7
11	206.7	62.4	59.0	62.8	61.8	111.8	61.7	61.9	61.7	63.0
12	21.6	20.3	20.1	20.5	20.0	21.8	19.2	19.3	21.1	21.5
13	22.1	33.5	32.4	18.9	21.8	33.3	18.1	18.9	33.5	14.5
14	33.3	22.3	22.4	25.3	33.7	23.3	28.9	33.8	21.7	15.2
15	15.7	15.3	18.2	33.4	17.7	18.4	15.5	22.7	15.6	28.0
16									170.8	171.2
17									21.6	21.3
Ref.	Barrero et al., 1999	Barrero et al., 1999	Lagnel et al., 2000	Grabley et al., 1996	Panasenko et al., 2004	Lagnel et al., 2000	Lu et al., 2009	Lu et al., 2009	Lagnel et al., 2000	Ramirez et al., 1993



C	<b>51</b> <sup>c</sup>	52 <sup>c</sup>	53 <sup>c</sup>	54 <sup>c</sup>	55 <sup>M</sup>	56 <sup>A</sup>	57 <sup>M</sup>	58 <sup>M</sup>	59 <sup>c</sup>	60 <sup>c</sup>
-	27.4	39.5	49.5	36.5	38.8	35.1	38.9	39.0	39.5	79.8
2	37.7	18.8	65.3	18.8	28.1	38.7	28.6	28.8	18.0	24.8
ß	78.9	42.0	51.7	42.7	79.5	215.1	79.5	79.7	41.7	39.6
4	38.8	33.0	35.4	33.1	39.8	47.9	39.8	39.9	33.0	32.8
v	49.4	49.4	50.5	54.1	50.7	51.8	50.7	50.8	48.9	48.5
9	23.3	23.6	24.4	24.3	24.3	24.4	24.2	24.5	25.2	27.8
٢	124.1	127.5	126.4	127.5	126.4	125.0	126.7	129.1	154.4	153.1
×	131.8	137.0	138.4	137.2	138.4	139.3	137.2	134.3	138.1	139.5
6	53.3	54.5	55.9	43.4	55.8	54.7	52.1	55.8	60.2	59.8
10	35.9	35.7	38.5	35.9	36.6	36.2	36.9	36.8	36.8	43.5
=	64.3	61.5	61.2	63.1	61.2	60.9	63.5	60.9	201.9	203.9
12	21.7	67.5	66.8	67.7	67.0	66.7	65.7	68.7	193.2	192.9
13	14.7	33.2	23.2	21.7	15.0	14.4,	15.1	15.0	33.1	32.7
14	15.4	21.9	33.8	22.0	15.9	22.5	15.9	15.0	21.9	22.0
15	28.2	14.6	15.9	33.0	28.7	25.7	28.0	28.2	15.1	9.3
16	173.5						172.9	173.0		
17	60.9						21.1	21.1		
Ref.	Zhao et al., 2014	Aasen et al., 1977	Xiao et al., 2017	Derita et al., 2013	Aranda et al., 1992	Zhao et al., 2014	He et al., 2014	He et al., 2014	Santoso et al., 2018	Dacunto, 2012



C	61 <sup>c</sup>	62 <sup>c</sup>	63 <sup>c</sup>	641	65 <sup>0</sup>	99 <sub>0</sub>	67 <sup>p</sup>	68 <sup>c</sup>	69c	70c
1	37.3	37.0	37.1	74.7	37.2	31.7	42.4	31.9	36.5	31.1
2	25.0	23.2	18.3	25.3	18.1	18.3	32.6	19.9	18.5	18.6
3	78.3	7.97	42.0	39.6	41.8	41.8	17.3	44.2	41.8	41.6
4	36.5	36.4	32.8	32.7	32.6	33.5	36.2	34.0	33.2	33.0
v	48.4	48.6	44.1	43.6	43.1	42.2	47.9	45.2	47.6	42.3
9	26.7	24.7	25.5	27.5	24.4	26.5	67.2	66.2	25.7	24.1
7	154.0	153.3	153.5	152.7	142.3	158.3	158.7	148.5	152.2	131.8
×	138.1	138.2	137.3	137.5	126.3	141.0	139.3	141.3	136.4	138.2
6	60.0	59.9	58.4	54.9	60.6	78.3	77.6	77.5	149.2	75.7
10	38.8	37.7	37.6	43.1	36.7	42.0	43.1	41.7	37.4	40.5
11	201.6	201.3	202.3	203.1	203.7	203.0	202.5	201.0	193.6	62.6
12	193.0	192.9	192.8	192.9	167.5	193.4	192.3	192.9	109.4	6.99
13	27.9	27.9	32.7	32.3	32.5	33.5	17.1	32.6	32.4	30.9
14	15.4	16.5	21.8	22.1	21.4	22.6	21.9	24.7	21.6	22.3
15	15.2	16.5	21.4	14.8	21.1	17.6	35.8	17.7	19.8	15.3
16		170.7						170.0		
17		21.2						21.3		
Ref.	Dacunto, 2012	Dacunto, 2012	Rodriguez et al., 2005	Dacunto, 2012	Liu et al., 2010	Mashimbye et al., 1999	Kioy et al., 1989	Mahmoud et al., 1980	Jansen, 1993	Barrero et al., 1999



C	м12	72 <sup>n</sup>	73 <sup>c</sup>	74	75 <sup>M</sup>	<u> 16</u> м	мLL	78 <sup>c</sup>	м62	80 <sup>M</sup>
1	33.6	32.2	39.4	40.1	38.8	38.9	38.8	39.1	38.6	38.7
2	19.8	18.2	18.6	18.6	28.8	28.8	28.7	39.1	27.8	27.7
3	45.6	43.1	41.9	42.0	81.4	81.3	81.3	35.5	73.7	73.5
4	35.1	32.8	32.8	33.2	43.1	43.1	43.0	35.6	43.6	43.6
S	47.5	45.7	49.4	49.5	51.7	51.6	51.1	42.8	43.0	43.0
9	65.5	77.1	23.3	24.8	24.4	24.4	24.5	23.6	24.0	24.0
7	131.0	125.1	126.4	151.0	126.2	126.7	128.8	127.3	126.3	126.7
æ	140.8	140.6	136.0	139.1	138.7	137.4	134.4	137.3	138.5	137.2
6	76.6	74.4	50.5	55.3	55.8	52.2	55.7	54.7	55.7	52.1
10	41.6	42.0	35.7	35.7	36.5	36.8	36.6	34.8	36.4	36.7
11	63.4	61.9	63.1	176.6	61.3	63.6	6.09	61.6	61.4	63.6
12	64.7	61.1	65.9	192.6	66.8	65.8	68.6	67.7	67.0	65.9
13	19.4	17.5	33.1	33.3	16.1	16.1	16.0	71.8	15.7	15.8
14	25.3	36.2	21.8	22.1	65.0	65.0	65.0	18.0	12.6	12.6
15	33.5	23.3	14.3	15.2	23.3	23.3	23.3	15.3	66.6	66.5
16		53.8,	170.7			173.0	173.0			173.0
17			21.0			21.2	21.1			21.2
Ref.	Grabley et al., 1996	Lu et al., 2009	Barrero et al., 1995	Fukuyama et al., 1985	He et al., 2014	He et al., 2014	He et al., 2014	Chaudhary et al., 2008	He et al., 2014	He et al., 2014



C	81 <sup>c</sup>	82 <sup>c</sup>	83c	84 <sup>c</sup>	85 <sup>c</sup>	86 <sup>c</sup>	87 <sup>c</sup>	88c	80c	90c
-	37.3	36.2	36.9	35.5	34.9	35.6	36.2	36.3	35.4	36.0
2	19.5	18.7	19.0	18.5	18.7	18.6	18.9	18.9	18.5	18.6
3	42.0	41.2	41.8	41.1	41.0	41.2	41.5	41.6	41.1	41.3
4	33.4	33.0	33.3	33.0	33.1	32.8	33.3	33.3	33.1	32.8
v	51.9	50.0	51.8	50.2	50.0	49.6	51.3	51.4	50.1	49.6
9	19.5	35.2	19.0	35.5	35.0	29.4	18.7	18.8	35.3	29.5
٢	34.1	199.9	33.8	201.2	201.0	75.1	31.3	33.8	200.4	72.8
×	124.3	129.3	132.5	132.4	134.7	136.4	136.0	135.3	134.5	136.8
6	136.2	165.3	141.1	162.8	169.5	150.5	146.2	135.5	158.2	138.7
10	38.5	40.4	38.1	39.9	40.2	39.3	38.2	38.0	40.0	38.9
11	19.8	14.9	58.4	58.5	58.1	57.9	57.9	60.7	60.0	60.4
12	19.8	11.9	21.7	11.3	56.5	61.1	64.0	21.3	11.5	21.5
13	33.4	21,2	33.3	21.2	21.1	21.5	20.4	33.3	32.5	33.0
14	21.8	32.5	20.8	32.5	32.4	33.0	33.2	21.6	21.2	21.1
15	19.5	17.5	19.4	18.3	18.0	20.0	21.6	19.5	18.3	15.0
16									170.7	171.0
17								20.7	20.9	20.5
Ref.	Yonemura et al., 2012	Panasenko et al., 2004	Kuchkova et al., 2005	Panasenko et al., 2004	Panasenko et al., 2004	Panasenko et al., 2004	Benites et al., 2001	Barrero et al., 1999	Barrero et al., 1999	Barrero et al., 1999



C	<b>91</b> <sup>c</sup>	92 <sup>c</sup>	93 <sup>c</sup>	94 <sup>c</sup>	95 <sup>c</sup>	96 <sup>c</sup>	97 <sup>c</sup>	98c	66	100
1	35.6	42.3	40.7	35.1	34.6	34.5	35.7	37.4	40.5	39.2
2	18.5	18.6	18.9	18.5	18.5	18.3	18.5	18.4	17.8	27.3
e.	41.1	42.9	43.5	41.1	42.0	41.3	41.2	42.7	42.6	78.3
4	32.9	33.1	33.4	33.2	32.7	32.9	33.3	33.5	32.2	38.3
S	42.2	53.2	54.0	41.7	40.0	43.3	41.9	47.6	65.0	64.6
Q	28.6	69.2	70.5	35.9	28.6	17.2	34.7	68.6	204.7	205.0
7	69.7	36.0	36.9	209.4	70.2	25.9	207.7	35.3	60.7	62.1
æ	135.3	142.9	129.2	65.8	64.8	65.6	66.5	62.4	61.4	62.5
6	145.0	147.7	140.9	72.4	71.0	72.1	73.2	69.6	55.2	55.8
10	39.4	40.1	36.9	37.9	37.9	37.3	37.7	38.1	37.2	37.6
11	59.2	190.7	57.8	12.3	13.9	61.4	58.5	58.1	60.5	60.6
12	61.2	21.1	21.9	12.3	16.8	66.0	12.4	20.1	21.5	21.8
13	21.5	36.0	36.2	20.6	21.2	21.5	32.5	35.0	33.2	28.5
14	32.9	21.2	22.3	32.4	33.5	33.6	20.7	22.4	21.6	15.6
15	20.4	17.7	18.7	17.0	17.7	16.4	17.1	18.2	18.7	19.0
16	170.8	169.3	169.7					169.8		
17	21.1	21.0	21.4					21.3		
18	171.4									
19	21.1									
Ref.	Panasenko et al., 2004	Lagnel et al., 2000	Lagnel et al., 2000	Panasenko et al., 2004	Panasenko et al., 2004	Benites et al., 2001	Barrero et al., 1999	Lagnel et al., 2000	De Bernardi et al., 1980	De Bernardi et al., 1980

С	1	2	3	4	5	9	7	×	6	10	11	12	13	14	15	16	17	18	19	
101 <sup>c</sup>	43.2	18.8	44.5	34.3	54.3	63.2	65.2	63.2	55.2	35.7	60.8	22.6	33.3	24.7	19.0					Lagnel et al., 2000
102 <sup>c</sup>	40.7	18.8	35.1	31.9	52.6	136.5	130.2	144.2	142.9	39.1	190.8	22.5	32.4	22.5	15.8					Lignel et al., 2000
103 <sup>c</sup>	32.1	18.4	38.6	32.8	51.2	115.6	129.0	139.7	146.8	40.5	59.7	11.9	34.4	22.5	15.7	169.3	20.4	170.7	20.8	Vlad et al., 2006
104 <sup>c</sup>	34.3	18.0	40.3	37.4	172.6	123.8	186.9	135.2	154.7	43.3	60.2	11.0	28.3	32.3	25.1	170.7	20.7			Vlad et al., 2006
105 <sup>c</sup>	33.9	17.9	40.0	37.5	173.1	124.1	185.3	133.3	160.4	43.8	59.4	57.2	28.6	31.7	25.4	170.3	20.6	170.8	20.9	Vlad et al., 2006
106 <sup>D</sup>	36.1	29.8	39.7	155.6	47.7	23.5	36.7	76.8	59.7	40.6	60.7	65.0	106.9	20.6	14.4					He et al., 2015
107 <sup>c</sup>	31.7	30.8	38.1	151.8	40.1	27.7	151.8	139.6	77.4	42.2	201.3	192.7	106.0	15.1	18.1					Mashimbye et al., 1999
108 <sup>c</sup>	39.4	31.6	38.5	151.3	45.8	27.0	152.9	137.9	58.3	38.2	201.1	193.3	106.0	18.4	13.5					Mashimbye et al., 1999
109	31.7	31.8	38.8	149.1	50.4	66.1	153.7	139.3	77.6	44.1	200.5	192.6	106.7	18.2	15.8					Wube et al., 2005
110 <sup>c</sup>	39.3	18.8	42.0	32.9	49.4	23.6	127.4	136.9	54.4	35.6	61.4	67.4	33.2	21.9	14.5					Derita et al., 2013



Ref.



C	111 <sup>c</sup>	112 <sup>A</sup>	113 <sup>c</sup>	114 <sup>c</sup>	115 <sup>c</sup>	116 <sup>A</sup>	117 <sup>c</sup>	118 <sup>c</sup>	119 <sup>M</sup>	120 <sup>M</sup>
1	38.5	31.3	37.6	39.8	33.4	49.9	30.5	47.4	25.1	25.9
2	18.3	25.8	27.1	18.5	18.1	64.0	17.8	66.0	25.7	25.2
3	42.4	75.5	79.0	42.4	35.8	52.4	44.7	55.2	76.7	78.7
4	33.1	37.9	38.8	32.9	38.0	35.0	34.1	37.2	38.9	39.3
S	49.7	43.6	49.2	49.8	50.8	50.2	45.6	50.6	37.6	41.4
6	23.4	23.5	23.5	23.6	23.2	24.2	65.4	66.2	24.8	66.4
٢	121.2	121.3	116.9	117.1	117.0	116.8	127.8	129.5	122.9	124.8
×	129.9	131.6	136.3	136.5	136.6	138.4	132.8	135.1	139.0	140.1
6	53.7	54.0	61.4	61.6	61.8	62.5	74.9	76.2	78.5	78.5
10	34.4	34.5	33.2	33.4	39.0	35.6	37.6	40.9	38.2	38.8
11	175.2	175.5	99.2	99.4	99.4	99.5	175.3	177.8	99.1	99.3
12	69.8	70.2	68.8	68.9	69.0	68.5	69.1	70.9	68.2	67.9
13	21.4	22.1	14.9	33.1	26.6	22.7	32.6	33.8	22.9	25.6
14	33.0	28.7	27.7	21.5	64.8	33.5	25.0	26.8	29.1	28.4
15	13.9	14.3	14.1	14.0	15.0	15.2	18.9	20.8	16.5	19.4
Ref.	Rukachai- sirikul et al., 2010	Kuang et al., 2016	Echeverri et al., 1997	Derita et al., 2013	Ayer and Craw, 1989	Kuang et al., 2016	Shiono et al., 2007	Shiono et al., 2007	Yang et al., 2013	Yang et al., 2013

Montagnac et al.,1996	Opiyo et al., 2011	Aranda et al., 2001	Aranua et al., 2001	narmance- naina et al., 2008	Au et al., 2009b	Opiyo, 2011	Chen et al., 2016	2005 ct al.,	Opiyo 2011	Ref.
				54.9						19
				56.6						18
		21.7		21.3				21.2		17
		171.3		170.1				170.7		16
33.5	14.97	21.0	20.9	18.0	21.3	21.5	14.7	13.5	13.1	15
21.6	22.5	32.9	28.2	24.2	33.0	33.3	28.3	27.6	21.0	14
20.3	32.3	21.5	15.6	32.9	16.1	17.0	15.4	15.9	32.5	13
172.0	170.2	171.4	174.7	104.1	168.7	169.3	167.6	169.7	169.8	12
99.1	67.7	97.2	68.2	104.3	98.5	74.3	99.5	6.9	6:99	11
37.0	36.8	40.8	36.4	38.9	39.2	38.5	34.4	34.1	32.8	10
167.7	171.7	164.9	170.0	76.7	75.7	77.3	59.7	50.6	49.4	6
128.3	122.3	129.7	123.8	131.5	129.2	130.0	129.7	127.2	127.0	×
21.5	131.7	21.6	21.6	134.9	143.3	141.1	135.5	135.7	135.9	7
18.0	117.6	17.7	18.1	67.3	25.2	25.3	25.2	24.7	24.7	9
51.5	52.3	45.2	50.6	44.9	41.1	41.7	49.6	49.4	50.5	S
33.5	32.7	33.4	39.0	33.6	33.0	32.7	39.4	37.7	33.9	4
41.7	40.5	35.1	78.3	44.5	41.1	31.3	78.3	80.2	41.9	θ
18.3	17.9	22.3	27.1	19.6	17.5	17.9	27.9	23.5	18.2	2
35.4	33.5	74.7	34.2	31.9	30.3	41.6	37.9	36.9	39.1	-
130 <sup>c</sup>	129 <sup>c</sup>	128 <sup>M</sup>	127 <sup>M</sup>	126 <sup>c</sup>	125 <sup>bC</sup>	124 <sup>c</sup>	123 <sup>A</sup>	122 <sup>c</sup>	121 <sup>c</sup>	υ









Table 3 Continued

151 <sup>c</sup>	39.6	18.6	42.3	33.3	53.2	22.6	23.3	114.3	64.5	37.1	98.3	134.6	33.7	21.8	14.0	172.9	30.3	14.1		Gaspar et al., 2008
150 <sup>M</sup>	48.2	64.8	51.0	34.6	52.7	130.4	129.4	79.0	70.2	40.9	101.7	80.9	23.0	33.4	15.8					Kuang et al., 2016
149 <sup>M</sup>	70.9	119.5	144.0	35.2	42.7	18.7	21.7	132.6	161.2	40.5	91.0	170.6	21.9	31.4	20.6	169.5	169.8	21.3	21.3	Aranda et al., 2001
148 <sup>M</sup>	71.8	119.2	144.0	35.2	42.7	18.6	21.3	130.6	163.3	40.0	97.3	171.5	22.2	31.3	19.5	171.6	21.3			Aranda et al., 2001
147 <sup>M</sup>	70.0	123.6	143.0	35.9	42.5	20.0	22.0	131.0	166.9	43.0	99.2	173.6	22.4	31.5	19.6					Aranda et al., 2001
146 <sup>M</sup>	71.5	118.9	144.6	35.0	42.2	18.8	21.3	126.7	165.4	39.7	68.5	174.0	22.0	31.2	21.1	170.5	21.6			Aranda et al., 2001
145 <sup>M</sup>	69.4	122.6	143.0	35.1	41.3	18.8	21.2	125.9	167.6	41.3	69.2	174.7	21.8	31.4	20.9					Aranda et al., 2001
144 <sup>M</sup>	36.2	119.8	138.7	34.6	48.2	18.9	21.5	124.1	168.5	35.5	68.3	174.1	22.2	32.7	20.6					Aranda et al., 2001
143 <sup>c</sup>	35.0	22.0	42.5	33.7	47.0	61.9	70.3	160.6	136.4	35.8	172.4	172.4	39.2	29.3	19.1					Mahmoud et al., 1980
142 <sup>c</sup>	36.3	20.6	43.0	35.4	50.6	69.69	66.6	150.6	140.3	33.3	171.0	70.2	33.2	22.9	18.3	171.1	21.3	169.8	20.8	Opiyo et al., 2011
141 <sup>c</sup>	33.1	20.8	43.1	35.4	49.3	69.8	73.8	154.5	137.9	36.5	172.1	66.1	33.4	23.1	18.4	170.9	21.4			Mahmoud et al., 1980
U	1	2	3	4	S	9	7	×	6	10	11	12	13	14	15	16	17	18	19	Ref.





υ	152 <sup>M</sup>	153 <sup>c</sup>	154 <sup>0</sup>	155 <sup>D</sup>	156 <sup>a</sup>	157 <sup>M</sup>	158 <sup>B</sup>	159 <sup>c</sup>	160 <sup>c</sup>	161 <sup>c</sup>	162 <sup>c</sup>
1	49.8	37.2	30.8	29.7	40.0	40.4	41.1	39.7	32.4	31.0	33.7
2	64.7	18.2	24.6	25.0	19.3	18.1	18.0	32.3	31.7	32.2	31.8
3	51.9	42.3	73.9	73.9	43.2	42.0	41.6	38.8	38.5	38.3	38.5
4	35.1	33.1	37.0	37.0	33.9	32.9	32.6	151.6	152.2	130.5	152.0
v	44.9	55.5	47.9	47.3	56.6	51.4	51.2	45.9	39.9	40.2	44.2
9	23.7	21.2	17.0	20.4	22.3	18.4	18.0	26.9	27.3	27.5	31.5
٢	59.6	28.7	33.9	27.8	29.0	22.4	32.0	135.5	139.8	141.4	62.3
*	65.2	38.3	77.6	37.8	40.4	37.4	76.0	127.0	129.9	130.5	156.9
6	62.3	57.4	58.1	56.0	66.3	49.9	65.0	49.3	76.9	76.5	137.4
10	36.0	35.7	35.2	35.0	35.5	35.4	37.2	36.7	40.6	53.7	36.8
11	100.1	175.8	174.8	175.7	99.2	67.6	100.0	67.9	75.0	98.6	172.1
12	68.0	71.2	75.8	70.6	71.4	179.1	174.0	170.2	169.6	167.3	69.9
13	23.4	33.5	16.4	15.3	34.1	14.5	33.4	105.5	105.7	105.9	104.7
14	33.3	21.2	21.6	21.6	21.8	22.0	21.8	18.6	15.9	18.5	18.1
15	16.2	15.5	29.1	29.1	15.6	33.5	16.4	12.5	18.5	15.2	16.4
Ref.	Kuang et al., 2016	Paul et al., 1997	He et al., 2015	He et al., 2015	Montegnac et al., 1996	Aranda et al., 2001	Sakio et al., 2001	Wube et al., 2005	Mashimbye et al., 1999	Rajab and Ndegwa, 2000	Wube et al., 2005



U	163 <sup>c</sup>	164 <sup>c</sup>	165 <sup>c</sup>	166 <sup>c</sup>	167 <sup>c</sup>	168 <sup>c</sup>	169 <sup>c</sup>	170 <sup>c</sup>	171 <sup>c</sup>	172 <sup>c</sup>
1	39.5	36.2	39.6	39.8	37.8	37.1	42.4	42.6	38.7	36.9
2	19.0	18.6	18.4	18.7	18.4	18.7	18.6	18.6	18.3	18.2
3	42.1	42.4	42.3	42.1	42.7	42.7	42.7	42.6	41.9	42.0
4	33.6	33.5	33.5	33.7	33.1	33.0	33.0	33.1	33.0	33.0
5	54.2	44.0	56.1	54.8	46.5	44.5	54.4	54.4	53.3	45.2
9	23.6	23.1	17.2	20.9	17.4	21.3	20.7	18.4	19.9	20.0
7	41.9	37.3	35.4	36.8	30.5	30.4	43.0	41.3	40.0	35.5
8	212.6	216.2	72.9	72.2	75.1	69.0	71.1	71.0	74.0	72.6
6	58.0	58.7	48.9	52.8	49.1	49.0	59.8	57.1	86.9	84.1
10	41.5	39.1	37.5	37.7	36.5	37.7	35.0	34.7	39.4	39.0
11	6.9	13.1	11.7	10.0	14.8	6.7	I	I	I	I
12	I	I	I	I	I	I	29.1	33.1	22.2	27.0
13	33.5	33.5	33.7	33.6	33.6	33.4	33.2	33.3	32.2	33.0
14	21.8	22.0	21.8	21.8	22.0	21.7	21.2	21.4	21.6	21.7
15	13.9	22.1	15.2	13.6	23.7	22.3	20.9	20.9	13.7	20.4
	Ohloff and	Wahlberg et	Wahlberg et	Wahlberg et	Wahlberg et					
Ref.	Giersch,	Giersch,	Giersch,	Giersch,	Giersch,	Giersch,	al., 1981	al., 1981	al., 1981	al., 1981
	C861	C861	C861	C861	C861	C861				



183 <sup>D</sup>	33.6	18.1	42.2	32.3	40.1	22.9	140.8	132.2	71.4	36.9		168.0	32.6	21.4	18.4			Liu et al.,	2010
182 <sup>c</sup>	32.8	18.1	41.5	33.6	49.0	24.2	145.1	135.8	206.7	45.2	1	62.4	32.3	22.2	17.1			Cuellar et	al., 2003
<b>181</b> <sup>c</sup>	32.6	17.9	41.4	33.7	48.4	25.2	156.2	189.9	203.4	45.3	1	189.9	32.2	22.1	16.8			Cuellar et	al., 2003
<b>180</b> <sup>c</sup>	33.2	18.2	41.6	33.6	49.4	24.4	143.4	132.9	205.9	45.1	1	16.4	32.3	22.2	17.2			Cuellar et	al., 2003
179 <sup>c</sup>	20.0	34.4	41.0	34.9	36.4	18.3	207.5	60.3	71.5	41.1	1	15.5	33.7	32.1	17.0			Montene-	gro et al., 2014
178 <sup>c</sup>	36.3	21.2	39.9	34.5	41.5	18.2	71.5	58.9	70.7	33.0	1	18.2	25.0	32.5	19.5	170.4	21.1	Montene-	gro et al., 2013
177 <sup>c</sup>	22.0	32.7	41.4	33.2	39.0	17.8	208.2	56.2	59.5	46.1	1	16.8	22.5	32.6	16.9			Cuellar et	al., 2003
176 <sup>c</sup>	38.0	18.5	41.1	32.9	50.3	35.6	202.3	133.9	158.9	18.3	1	61.4	32.2	20.9	36.7			Cuellar et	al., 2003
175 <sup>c</sup>	38.3	18.1	40.8	33.0	50.4	34.0	200.8	137.7	163.1	38.3	1	193.8	32.4	21.0	18.1			Bastos et	al., 1999
174 <sup>c</sup>	38.0	18.5	41.5	33.0	48.6	27.0	67.7	139.1	164.5	37.7	1	196.2	32.7	21.3	20.1			Fukuyama	et al.,1985
173 <sup>c</sup>	38.4	18.6	41.2	32.8	50.6	35.4	201.6	131.2	158.3	36.9	1	15.5	32.2	20.9	18.5			Cuellar et	al., 2003
U	-	7	3	4	S	9	7	8	6	10	11	12	13	14	15	16	17	Ref	

\*Solvent are indicated by superscripts A (Acetone-d6), B (C6D6), C (CDCl3), D (DMSO-d6), DC (CD2Cl2), P (C5D5N), M (CD5OD). a, b, c within a vertical column may be reversed.



### 2.2. Drim-8(12)-ene sesquiterpenes

Compounds 25-34 (Table 4) are examples of some previously reported drimane sesquiterpenes. Generally, their olefinic carbons C-8 and C-12 resonate between  $\delta$  143.1-161.5 and  $\delta$  106.3-109.9 ppm, respectively (Aasen et al., 1977; Barrero et al., 1995; Kinoshita et al., 2002; Justicia et al., 2005; Delgado et al., 2008; Xu et al., 2009a). In drim-8(12)-enes with no substituents in close proximity to the 8:12 double bond, the chemical shifts of the olefinic C-8 and C-12 carbons are ~151 and ~101 ppm, respectively. However, the presence of an additional double bond (conjugation) has a pronounced effect on the olefinic carbons resonances. For example, the olefinic carbons C-8 and C-12 in drimanes such as drim-8(12),9(11)-diene (26) resonate more downfield at & 161.5 and 108.8 ppm, respectively, i.e. the olefinic carbons are deshielded by 10.0 ppm

#### Table 4

Structures of drimanes 25-34.

and 7.3 ppm for C-8 and C-12, respectively (Table 3). The additional olefinic carbon atoms in compound 26 resonate at  $\delta$  149.9 and 103.0 ppm for C-9 and C-11, respectively (Kinoshita et al., 2002). The presence of a hydroxyl group at C-11 cause characteristic downfield (~5 ppm) and upfield shifts (~ 4 ppm) for the olefinic C-I2 and C-8, respectively. In the presence of two hydroxyl groups (at C-7 and C-11), the olefinic carbon C-12 is further deshielded, while C-8 becomes shielded (cf. drimanes 25, 27 and 28). Acetylation of the hydroxyl group at C-11 further deshields C-12 but shields C-8 as evidenced by <sup>13</sup>C shifts of drimanes 27 and 30. Mono and di-acetylated derivatives of 8(12)-drimenes such as 29-34 having the acetyl group mostly at C-3 or C-11 were reported (Barrero et al., 1995; Toshima et al., 2001; Justicia et al., 2005; Dacunto, 2012; Derita et al., 2013).

	<b>1</b> <b>1</b> <b>1</b> <b>1</b> <b>1</b> <b>1</b> <b>1</b> <b>1</b> <b>1</b> <b>1</b>	
<b>26</b> Δ <sup>9(11)</sup>	<b>29</b> 3β-OAc, 11-OH	<b>32</b> 3β OH, 11-OAc
27 11-OH, albicanol	<b>30</b> 11-OAc, albicanylacetate	<b>33</b> 3β, 11-OAc
<b>28</b> 7α, 11-ΟΗ	<b>31</b> 7α-OH, 11-OAc	<b>34</b> 3β-OAc, 11- CO <sub>2</sub> Me

11

#### 2.3. Drim-7-ene sesquiterpenes

<sup>13</sup>C NMR data of several drim-7-enes have been reported and compounds 35-80 (Table 5) are some of the most important examples (Panasenko et al., 2004; Lu et al., 2009; Xu et al., 2009a; Dacunto, 2012; Yonemura et al., 2012; Derita et al., 2013; Zhao et al., 2014; Xiao et al., 2017; Santoso et al., 2018). Hydroxylation is mostly observed at C-11, C-9, C-6 (Aasen et al.,1977; Barrero et al.,1995; Grabley et al.,1996 Chaudhary et al., 2008; Lu et al., 2009; He et al., 2014; Xiao et al., 2017). Drim-7-ene derivatives having the hydroxyl group at C-3 include compounds 38, 47, 50, 51, 55, 57-58, 61 and 62. In such compounds, the oxymethine C-3 carbon resonates between  $\delta$  78.9 and 79.7 ppm (Lu et al., 2009; Xu et al., 2009a; Dacunto, 2012; He et al., 2014; Zhao et al., 2014). Drim-7-ene derivatives having hydroxyl group at C-9 include 42-45, 47, 48, 70-72 and the oxygenated C-9 carbon resonates between  $\delta$ 

74.4-76.6 ppm (Lagnel et al., 2000; Panasenko et al., 2004; Lu et al., 2009). Drim-7-ene derivatives having hydroxyl group at C-6 such as **43**, **44**, **67** and **71** have also been reported and their oxymethine C-6 carbon resonates between  $\delta$  65.4-68.4 ppm (Lagnel et al., 2000).

The dial derivatives **59-64** (Aasen et al., 1977; Rodriguez et al., 2005; Dacunto, 2012) and mono-al derivatives **41**, **65**, **69** and **74** are known (Fukuyama et al.1985; Jansen, 1993; Liu et al., 2010; Derita et al., 2013). The carbonyl carbons peaks appear between  $\delta$ 201.3 -203.9 and 192.3-193.9 ppm for C-11 and C-12 carbonyl carbons, respectively (Kioy et al., 1989; Rodriguez et al., 2005; Dacunto, 2012; Santoso et al., 2018). Changweikangic acid B (**65**), a drim-7-ene derivative with COOH substitution at C-12 had the carbonyl carbon resonating at  $\delta$  167.5 ppm (Liu et al., 2010), while polygonic acid (**74**) with COOH substitution at C11 had



the carbonyl carbon peak at  $\delta$  176.6 ppm (Fukuyama et al., 1985). Compound 65 was isolated from a mixture of *Daphniphyllum calycinum* and *Polygonum hydropiper*. Acetylated derivatives **49-51**, **57**, **58**, **63**, **68**, **73** 

and **80** have been reported with acetylation mainly occurring at C-11 (Mahmoud et al., 1980; Ramirez et al., 1993; Barrero et al., 1995; Lagnel et al., 2000; Rodriguez et al., 2005; He et al., 2014).

# Table 5

Structures of drimanes 35-80.



In drim-7-enes with no substitution in close proximity to the double bond, e.g. in drimane **40**, the C-7 and C-8 olefinic carbons resonate at  $\delta$  121.9 and 135.3 ppm, respectively (Yonemura et al., 2012). The presence of aldehyde functional group at C-11 in drimane **41** leads to downfield (~3.4 ppm) and upfield (~4.5 ppm) shifts for the olefinic C-7 and C-8, respectively. The presence of two aldehyde groups, at C-11 and C-12, leads to downfield shifts of ~32.5 and ~2.8 ppm for the olefinic C-7 and C-8, respectively (cf. drimanes **40** and **59**, Table 3). The presence of hydroxyl group at C-11 deshields C-7 by 2.0 ppm and shields C-8 by 1.9 ppm. It also leads to downfield (+49.7 ppm) and upfield (-5.0 ppm) shifts for C-11 and C-9, respectively (cf. drimanes **35** and **40**). In drim-7-enes such as **42** that have two hydroxyl groups (at C-9 and C-11), the olefinic C-7 and C-8 carbons resonate at ~127 and 135 ppm, respectively (Barrero et al., 1999). The presence



of hydroxyl group at C-3 in drim-7-enes such as in compound 38 deshields C-3 and C-2 by ~37 ppm and ~9 ppm, respectively, while C-1 is shielded by 0.5 ppm. The 3-oxo derivatives of drim-7-ene including 3-oxodrim-7-ene-11-ol (37) and 3-oxodrim-7-ene-11, 12-diol (56) were reported having C-3 carbonyl carbon resonating at  $\delta$  215.1-216.7 (Xu et al., 2009a; Zhao et al., 2014)]. The 6-oxo derivatives of drim-7-ene such as 39 and 45-40 have the carbonyl carbon resonating at ~200 ppm (Lagnel et al., 2000; Panasenko et al., 2004; Lu et al., 2009). The presence of the oxo group at C-6 deshields C-7 by ~6 and C-8 by ~22 ppm, thus the olefinic carbons resonate at about  $\delta$  129 and 156 ppm, respectively. Drim-7-enes such as 37 and 56 that have oxo group at C-3 are deshielded at C-3 and C-2 but shielded at C-1 by about 174, 20 and 5 ppm, respectively.

Drimenol (35), isodrimenol (36), polygodiol (52), isodrimenediol (54), polygodial (59) and isopolygodial (63) are sets on epimers at the C-9 stereogenic center. The configuration of their C-11 substituent is evidenced by the noticeable difference in the chemical shifts of the C-1, C-5 and C-15 $\gamma$  carbons (Rodriguez et

# Table 6

Structures of drimanes 81-93.

al 2005). For example, in polygodial (**59**) and isopolygodial (**63**), the difference in the chemical shifts of the C-1, C-5 and C-15 $\gamma$  carbons is observed [ $\Delta\delta = \delta$  (**59**) - $\delta$ (**63**): +2.4, +4.8 and -6.3 ppm, respectively].

### 2.4. Drim-8(9)-ene sesquiterpenes

Drimane derivatives **81-93** (Table 6) having a double at 8:9 have been reported by several workers, showing C-8 and C-9 olefinic carbons resonating between  $\delta$ 124.3-134.9 and 136.2-169.5 ppm, respectively (Benites et al., 2001; Kuchkova et al., 2005). Drim-8(9)-enes with no substituent in close proximity to the double bond, e.g. **81** have the olefinic C-7 and C-8 resonating at ~124 and ~136 ppm, respectively (Yonemura et al., 2012). Compounds **83-87** and **93** have hydroxyl group at C-11 and the oxymethylene carbon resonates between  $\delta$  57.7-58.5 ppm (Benites et al., 2001; Panasenko et al., 2004; Kuchkova et al., 2005). Compounds **85-87** have hydroxyl group at C-12 and the oxymethylene carbon resonates between  $\delta$  56.5 - 64.0 ppm (Benites et al., 2001; Kuchkova et al., 2005).

	1 $4$ $10$ $81$ $81$ $Drim-8(9)-ene$	
<b>82</b> 7-oxo	<b>86</b> 7β, 11, 12-OH	<b>90</b> 7β-OH, 11-OAc
<b>83</b> 11-OH	<b>87</b> 11, 12-OH	<b>91</b> 7β-OH, 11, 12-OAc
<b>84</b> 7-oxo, 11-OH	<b>88</b> 11-OAc	<b>92</b> 6-OAc, 11-CHO
<b>85</b> 7-oxo, 11, 12-OH	<b>89</b> 7-oxo, 11-OAc	<b>93</b> 6-OAc, 11-OH

The presence of hydroxyl group at C-11 deshields the olefinic carbons C-8 and C-9 by ~8 ppm and ~6 ppm, respectively. For example, in drim-8(9)-en-11-ol (83), C-8 and C-9 carbons resonate at  $\delta$  132.5 and 141.1 ppm, respectively (Kuchkova et al., 2005). In drim-8(9)-enes having two hydroxyl groups (at C-11 and C-12), the olefinic carbons are less shielded and a downfield shift of ~12 ppm and ~10 ppm are observed for C-8 and C-9, respectively. For example, in 11,12-dihydroxydrim-8-ene (87) the olefinic carbons C-8 and C-9 resonate at δ 136.0 and 146.2 ppm, respectively (Benites et al., 2001). Monoacetyl derivatives 88-90, 92, 93 and diacetyl derivatives such as 91 were also reported (Barrero et al., 1995; Lagnel et al., 2000; Panasenko et al., 2004). Acetylation of the hydroxyl group at C-11 further deshields C-8 but shields C-9 by ~2.8 and ~5.6 ppm. In 11-acetoxy-8-drimene (88), the olefinic carbons C-8 and C-9 resonate at  $\delta$  135.3 and 135.5 ppm,

respectively, while in 11-acetoxy-8-drimen-7 $\beta$ -ol (90), they resonate at 136.8 and 138.7 ppm, respectively (Barrero et al., 1999). The 7-oxo derivatives of drim-8(9)-ene such as 82, 84, 85 and 89 have the C-7 resonating at  $\delta$  199-202 ppm (Benites et al., 2001; Kuchkova et al., 2005). The presence of oxo group at C-7 deshields the olefinic carbons C-8 and C-9 by ~5-10 and ~22-44 ppm, respectively, making the olefinic carbon to resonate between  $\delta$  129-137 and 158-170 ppm, respectively. In drim-8-en-7-one (83), C-7, C-8, and C-9 resonate at  $\delta$  199.9, 129.3 and 165.3, respectively (Panasenko et al., 2004).

2.5.  $8\alpha$ ,  $9\alpha$  and  $7\alpha$ ,  $8\alpha$ -Epoxydrimane sesquiterpenes <sup>13</sup>C NMR chemical shifts of synthetic  $8\alpha$ , $9\alpha$ -epoxydrimanes (Table 7) have been reported by several groups of workers. The <sup>13</sup>C values of some representatives **94-98** are shown in Table 3. The oxygenated carbons C-8 and C-9 resonate



between  $\delta$  64.8-66.5 and 69.6-73.2 ppm, respectively (Lagnel et al., 2000; Benites et al., 2001; Panasenko et al., 2004). In 7-oxo-8 $\alpha$ ,9 $\alpha$ -epoxydrimanes derivative, e.g. 11-hy-droxy-8 $\alpha$ ,9 $\alpha$ -epoxydriman-7-one (97), C-7, C-8 and C-9 resonate at  $\delta$  207.7, 66.5 and 65.5 ppm, respectively (Barrero et al., 1999). Hydroxylated derivatives **95-98** having the hydroxyl group mostly at C-7, C-11 and C-12 have been reported (Barrero et al., 2004). In the synthetic 7 $\alpha$ , 8 $\alpha$ -epoxydrimane derivatives, e.g. **99-101**, the C-7 and C-8 oxygenated carbons resonate at  $\delta$  60.7 and 65.2 ppm (De Bernardi et al., 1980; Lagnel et al., 2000). The 6-oxo derivatives (**99** and **100**) had the C-6 carbon resonate between  $\delta$  204.5 and 205.0 ppm (De Bernardi et al., 1980; Lagnel et al., 2000).

### Table 7

Structures of drimanes 94-105.

2.6. Drim-6,8-diene and drim-5,8-diene sesquiterpenes

13C signals of drim-6,8-diene and drim-5,8-diene sesquiterpenes such as **102-103** and **104-105**, respectively (Table 7) were reported. In drim-6,8-dien-11-al (**102**) and 7,11-diacetoxydrima-6,8-diene (**103**), the olefinic carbons C-6, C-7, C-8 and C-9 resonate at  $\delta$  136.5, 130.2, 144.2, 142.9 and at 115.6, 129.0, 139.7 and 146.8 ppm, respectively (Lagnel et al., 2000; Vlad et al., 2006). In 6-oxodrim-5,8-diene derivatives **104** and **105**, the olefinic carbons C-5, C-9, C-8 and C-6 resonated between  $\delta$  172.6-173.1, 154.7-160.4, 135.2-133.3 and 123.9-124.1 ppm, respectively, while the C-7 carbonyl carbon in the compounds resonated between  $\delta$  186.9-185.3 ppm (Vlad et al., 2006).



#### 2.7. Rearranged drimane sesquiterpenes

Assignment of <sup>13</sup>C signals of a number of rearranged drimane sesquiterpenes, e.g. **106-109** (Table 8) have been reported (Wube et al., 2005; Opiyo, 2011; He et al., 2015). Sulphureine D (**106**), muzigadial (**107**), 9-deoxymuzigadial (108) and  $6\alpha$ -hydroxymuzigadial (**109**) have an exocyclic double bond at C-4 and a methyl group at C-3. The olefinic carbons C-4 and C-13 resonate between  $\delta$  149.1-155.6 and 106.0 -106.9 ppm, respectively (Wube et al., 2005; He et al., 2015). In 106, the hydroxylated carbons C-11 and C-12 resonate at  $\delta$  60.7 and 65.0 ppm, respectively (Table 3). In drim-4(13)-ene derivatives **107-109** which have additional double bond at 7:8, olefinic carbons C-7 and C-8 resonate between  $\delta$  150.8-153.8 and 139.6-139.3 ppm, respectively (Mashimbye et al., 1999; Wube et al., 2005).

### 2.8. Tricyclic drim-7-ene sesquiterpenes

<sup>13</sup>C NMR assignments of some tricyclic drim-7-enes **110-126** (Table 8) are shown in Table 3 (Wube et al., 2005; Shiono et al., 2007; Harinantenaina et al., 2008; Xu et al., 2009b; Opiyo, 2011; Derita et al., 2013; Chen et al., 2016; Kuang et al., 2016). In dehydroxydrimeninol (**110**), the olefinic carbons C-7 and C-8 resonate at  $\delta$  127.4 and 136.9 ppm, respectively, while the oxymethylene carbons C-11 and C-12 resonate at  $\delta$  61.4 and 67.4 ppm, respectively (Derita et al., 2013). The presence of hydroxyl group at C-11 deshields C-11 (~38 ppm) and C-12 (~ 1.5 ppm) but shields C-7 (~ 10 ppm) and C-8 (~0.4

ppm). For example, in isodrimeninol (**114**), C-7, C-8, C-11 and C-12 resonate at  $\delta$  117.1, 136.5, 99.4 and 68.9 ppm, respectively (Derita et al., 2013).

The lactone carbonyl carbon of 11,12 olides such as compounds 111, 112, 117 and 118 resonates between  $\delta$  175-178 ppm (Shiono et al., 2007; Rukachaisirikulet al., 2010; Kuang et al., 2016). The presence C-11 ester group, e.g. drimenin (111) shields the C-7 and C-8 olefinic carbons by 6.2 and 7.0 ppm, respectively but deshields the oxymethylene C-12 carbon by 2.4 ppm. In compound 111, C-7, C-8 and C-12 <sup>13</sup>C peaks were observed at δ 121.2, 129.9 and 69.8 ppm, respectively (Rukachaisirikul et al., 2010). In 12,11 olides such as compounds 121-123, the carbonyl carbon resonates more upfield between  $\delta$  167-171 ppm as a result of conjugation. The presence of C-12 ester group, as in cinnamolide (121), deshields the olefinic C-7 carbon by 8.5 ppm but shields C-8 by 9.9 ppm. In drimanes such as 124-126, the oxygenated guaternary C-9 carbon resonates between  $\delta$  75.7-77.3 ppm (Harinantenaina et al., 2008; Xu et al., 2009b; Opiyo, 2011). The presence of the hydroxyl group at C-9 deshields C-7, C-8, C-9 and C-11 but shields C-12. For example, in 9α-hydroxycinnamolide (124), C-7, C-8, C-9 and C-11 resonate at δ 141.1 (+5.2), 130.0 (+2.8), 77.3 (+10.4) and 74.3 (+7.4) ppm, respectively. Compounds 124 and 125 were isolated form Warburgia ugandensis Engl. (Opiyo, 2011; Xu et al., 2009b), while cinnamodial 11α, 12β-dimethyl acetal (126) was isolated form Cinnamosma madagascariensis Danguy (Harinantenaina et al., 2008).



# Table 8

Structures of drimanes 106-126.

11	110-126
106-109	
<b>106</b> 3β-H, 8α, 11, 12-OH, sulphureuine D	<b>117</b> 6β, 9α-OH, 11-oxo, stobilactone A
<b>107</b> 3α-Η, 9α-ΟΗ, 11, 12-CHO, muzigadial	<b>118</b> 2α, 6β, 9α-OH, 11-oxo, stobilactone B
<b>108</b> 3α-Η, 11, 12-CHO, 9-deoxymuzigadial	<b>119</b> 3α, 9α, 11α –ΟΗ
<b>109</b> 3α-Η, 6α, 9α-ΟΗ, 11, 12-CHO	<b>120</b> 3α, 6β, 9α, 11α-ΟΗ
110 dehydroxydrimeninol	<b>121</b> 12-oxo, cinnamolide
<b>111</b> 11-oxo, drimenin	<b>122</b> 3β- OAc, 12-oxo
112 3α-ΟΗ 11-οχο	<b>123</b> 3 $\beta$ , 11 $\alpha$ -OH, 12-oxo, hydroxydendocarbin A
<b>113</b> 3β, 11α-OH, danilol	<b>124</b> 9α-OH, 12-οχο
<b>114</b> 11α-OH, isodrimeninol	<b>125</b> 9α, 11α-OH, 12-oxo, ugandenial A
<b>115</b> 11α, 14-ΟΗ	<b>126</b> 6β-OAc, 9α-OH, 11α, 12β-OMe
<b>116</b> 2α, 11α-ΟΗ	

2.9. Tricyclic drim-8(9)-ene lactone derivatives

In tricyclic drim-8(9)-ene sesquiterpenes such as 127-143 (Table 9), the lactone carbonyl carbon resonates between  $\delta$  164.9-172.4 ppm, while the olefinic carbons C-8 and C-9 resonate between δ 122.3-140.7 and 150.8 -171.7 ppm, respectively (Aranda et al., 1992; Montagnac et al., 1996; Sakio et al., 2001; Opiyo et al., 2011; Sultana et al., 2011). In drim-8(9)-ene lactones with no substituent in close proximity to the lactone ring, e.g. 3β-hydroxyconfertifolin (127), the olefinic carbons C-8 and C-9 resonate at  $\delta$  123.8 and 170.0, ppm respectively. Bemadienolide (129) has an additional double bond at C-6 evidenced by chemical shifts at  $\delta$  117.6 and 131.7 ppm for C-6 and C-7, respectively (Kioy et al., 1990; Opiyo et al., 2011). The presence of additional double bond shielded C-8 (1.5 ppm), C-11 (0.5 ppm), C-12 (4.5) but shielded C-9 (1.7 ppm) in compound 129 (Table 3). Compounds 130-132, 134 and 135, which have a hydroxyl group at C-11 showed the oxygenated C-11 carbon peak between  $\delta$  96.8-99.9 ppm which is equivalent to a downfield shift of ~33 ppm. The presence of hydroxyl group at C-11 deshields C-8 but shields C-9 and C-12. For example, in 11β-hydroxyconfertifolin (130), C-8, C-9 and C-12 peaks were observed at  $\delta$  128.3, 167.7 and 172.0 ppm, respectively (Montagnac et al., 1996). Compounds 131-134 have hydroxyl group at C-7 and the oxymethine carbon resonates between  $\delta$ 59.7-65.0 ppm which is more downfield by ~43 ppm. In dihydroxy-confertifolin (136) and diacetoxyconfertifolin (136), C-8, C-9, C-11 and C-12 carbons resonate at  $\delta$  130.6, 166.3, 99.9, 171.7 and 132.0, 162.9, 90.8, 170.5 ppm, respectively (Aranda et al., 2001). Winterin (137) was isolated from Polygonum hydropiper (Linn.) and its C-8, C-9, C-11 and C-12 carbons resonate at  $\delta$  140.7, 150.8, 170.6 and 169.9 ppm, respectively (Sultana et al., 2011).

2.10. Tricyclic drim-2,8-diene lactone sesquiterpenes

Synthetic drimane 12,11 olides such as **144-149** (Table 9) having two double bonds at C-2 and C-8 have been reported (Table 3). Their lactone carbonyl carbon resonates between  $\delta$ 170.6-174.7 ppm, while the olefinic carbons resonate between  $\delta$  118.9-123.6, 138.7-144.6, 124.1-132.6 and 161.2-168.5 ppm for C-2, C-3, C-8 and C-9, respectively (Aranda et al., 1992). For non-substituted compounds in this series, e.g.



2,3-dehyddroconfertifolin (**144**), the olefinic carbons C-2, C-3, C-8 and C-9 resonate at  $\delta$  119.8, 138.7, 124.1 and 168.5 ppm, respectively. Presence of hydroxyl group at C-1 deshields C-1, C-2, C-3, C-8, C-11 and C-12 but shields C-9. For example, in 1 $\alpha$ -hydroxy-2,3-dehyddroconfertifolin (**145**),C-1, C-2, C-3, C-8, C-9, C-11 and C-12 peaks were observed at  $\delta$  69.4

### Table 9

Structures of drimanes 127-149.

	11 9 7 7 7 7 7 7 7 7 7 7 7 7 7
<b>127</b> 3β-OH	<b>139</b> 11-οχο, 12β-ΟΗ
<b>128</b> 1α- ΟΑς, 11α-ΟΗ	<b>140</b> 6β, 7α-OH, 11-oxo, deacetoxyugandensolide
<b>129</b> $\Delta^6$ , bemadienolide	<b>141</b> 6β-OAc, 7α-OH, 11-oxo, ugandensolide
<b>130</b> 11β-OH	<b>142</b> 6β-OAc, 7α- OAc, 11-οχο
<b>131</b> 7β, 11α-OH, 12-oxo, fuegin	<b>143</b> 7α-OH, 11-oxo, futronolide
<b>132</b> 7α-OH, 11β-OEt, 12-oxo, dendocarbin J	<b>144</b> 12-οχο, Δ <sup>2</sup>
<b>133</b> 7α-OH, 11α-OEt, 12-oxo, dendocarbin K	<b>145</b> 1α-OH, 12-oxo, Δ <sup>2</sup>
<b>134</b> 7α, 11β-OH, 12-oxo, dendocarbin L	<b>146</b> 1α-OAc, 12-oxo, Δ <sup>2</sup>
<b>135</b> 1α, 11α-OH, 12-οχο	<b>147</b> 1α, 11α-ΟΗ, 12-οχο, Δ <sup>2</sup>
<b>136</b> 1α, 11α-OAc, 12-οχο	<b>148</b> 1α-OAc, 11α -OH, 12-oxo, Δ <sup>2</sup>
<b>137</b> 11, 12-oxo, winterin	<b>149</b> 1α, 11α -OAc, 12-oxo, Δ <sup>2</sup>
138 11-oxo, sodrimenin	

2.11. Miscellaneous unsaturated tricyclic drimane sesquiterpenes

In 11,12-epoxy-2 $\alpha$ ,8 $\alpha$ ,11 $\alpha$ -trihydroxydrim-6-ene (**150**) olefinic C-6 and C-7 carbon resonate at  $\delta$  130.4 and 129.4 ppm, respectively while the oxygenated carbon C-2, C-8, C-11 and C-12 peaks were observed at  $\delta$  64.8, 79.0, 101.7 and 80.9 ppm, respectively (Kuang et al., 2016). Drim-8(12)-ene 11,12-epoxide (**151**) showed the olefinic carbon peaks at  $\delta$  114.3 and 134.6 ppm acetal C-11 carbon peak  $\delta$  98.3 ppm (Gaspar et al., 2008). In 2 $\alpha$ -hydroxy-7 $\alpha$ ,8 $\alpha$ -epoxyisodrimeninol (**152**), the epoxide carbon peaks at  $\delta$  59.6 and 65.2 ppm for C-7 and C-8, respectively (Kuang et al., 2016). Compounds

**151** and **152** were isolated from endophytic fungi (Kuang et al., 2016).

(+33.2), 122.6 (+2.8), 143.0 (+4.3), 125.9 (+1.8), 167.6 (-0.9) ,

69.2 (+0.9) and 174.7 (+0.6) ppm, respectively (Aranda et al.,

2001). In derivatives 146, 148 and 149 having acetyl groups at

C-1, the oxymethine carbon C-1 resonate between 70.9 -71.8

ppm (Aranda et al., 1992).

2.12. Saturated tricyclic drimane sesquiterpenes 153-158

<sup>13</sup>C NMR signals for dihydrodrimenin (**153**), sulphureuine E (**154**), sulphureuine F (**155**) and dihydroisodrimeninol (**156**) were reported (Table 3). In dihydrodrimenin (**153**), the lactone carbonyl C-11 carbon peak was observed at  $\delta$  175.8 ppm, while C-8 and C-12 peaks were at  $\delta$  38.3 and 71.2 ppm, respectively. In sulphureuine F (**155**), C-8, C-9, C-11 and C-12 signals were observed at  $\delta$  37.8, 56.0, 175.7 and 70.6 ppm, respectively. The presence of hydroxyl group at C-8 deshielded



C-8, C-9, and C-12 but shielded C-11. For example, in sulphureuine E (**154**), C-8, C-9, C-11 and C-12 peaks were observed at 77.6 (+39.8), 58.1 (+2.1), 174.8 (-0.9) and 75.8 (+5.2) ppm, respectively. Drimanes **154** and **155** have hydroxyl group at C-3 and the oxymethine peak was observed at  $\delta$  73.9 ppm. Compounds **157** and **158** are drimane 12,11-lactone derivatives and their lactone carbonyl carbon resonated at  $\delta$  174.0-179.1 ppm (Table 3). Drimanes **153-156** were isolated from cultures of fungi.

2.13. Rearranged tricyclic drimane sesquiterpenes

### Table 10

Structures of drimanes 150-162.

Drim-4(13)-7-diene 12,11 olide derivatives **159-161** (Table 10) have been reported and their lactone carbonyl carbon resonates between  $\delta$  167.3-170.2 ppm (Mashimbye et al., 1999; Rajab and Ndegwa, 2000; Wube et al., 2005). In drim-4(13), 8-diene 11,12 olide derivative **162**, the carbonyl carbon peak was more downfield at  $\delta$  172.1 ppm. The olefinic carbons C-4, C-8, C-9 and C-13 resonate between  $\delta$  152.0, 156.9, 137.4 and 104.7 ppm, respectively (Table 3). Compounds **159, 161** and **162** were isolated from Warburgia ugandensis Sprague, while compound **160** was isolated from *Warburgia salutaris* (Bertol. f.) Chiov.

150-158	159-162
<b>150</b> 2α, 8α, 12α-ΟΗ, Δ <sup>6</sup>	157 12-oxo, confertifolin dihydro
<b>151</b> 12α-EtCOO, Δ <sup>8(12)</sup>	<b>158</b> 8α, 11α-OH, 12-oxo, dendocarbin D
<b>152</b> 7,8-ероху, 2α, 12α-ΟΗ	<b>159</b> 12-οχο, Δ <sup>7</sup>
<b>153</b> 11-oxo	<b>160</b> 9 $\alpha$ -OH, 12-oxo, $\Delta^7$ , muzigadiolide
<b>154</b> 3α, 8β-OH, 11-oxo, sulphureuine E	<b>161</b> 9α, 11α -OH, 12-οχο, Δ <sup>7</sup> ,
<b>155</b> 3α-OH, 11-oxo, sulphureuine F	<b>162</b> 7β-OH, 11-oxo, Δ <sup>8(9)</sup>
<b>156</b> 11α-ΟΗ	

2.14. Nor-drimane sesquiterpenes

Compounds belonging to the nordrimane group have been identified in *Capsicodendron dinisii* (Schwacke) (Bastos et al., 1999), *Polygonum hydropiper* (Fukuyama et al., 1985) and certain tobacco varieties (Wahlberg et al., 1981). The compounds are partly responsible for the characteristic flavor in tobacco. Some examples of 12-nor (**163-168**) and 11-nor (**169-18**) **3** drimane derivatives are given in Table 11. Their chemical shifts are summarized in Table 3 (Wahlberg et al., 1981; Ohloff and Giersch, 1985; Bastos et al., 1999; Cuellar et al., 2003; Liu et al., 2010; Montenegro et al., 2014).



# Table 11

Structures of nordrimane 163-183.

<b>163</b> 8-οχο, 11β -CH <sub>3</sub>	<b>174</b> 7β-OH, 12-CHO, Δ <sup>8(9)</sup> , isopolygonal	
<b>164</b> 8-oxo, 11α-CH <sub>3</sub>	<b>175</b> 7-oxo, 12-CHO, Δ <sup>8(9)</sup> , polygonone	
<b>165</b> 8β-OH, 11β -CH <sub>3</sub>	<b>176</b> 7-oxo, 12-OH, Δ <sup>8(9)</sup>	
<b>166</b> 8 α-OH, 11β -CH <sub>3</sub>	<b>177</b> 7-οχο, 8β, 9β-ероχу, 12α-CH <sub>3</sub>	
<b>167</b> 8β-OH, 11α-CH <sub>3</sub>	<b>178</b> 7β-OAc, 8α,9α- ероху, 12β -CH <sub>3</sub>	
<b>168</b> 8α -OH, 11α-CH <sub>3</sub>	<b>179</b> 7-οχο, 8α,9α- ероχу, 12β -CH <sub>3</sub>	
<b>169</b> 8α-OH, 12β -CH <sub>3</sub>	<b>180</b> 9-oxo, 12 -CH <sub>3</sub> , Δ <sup>7</sup>	
<b>170</b> 8β-OH, 12α-CH <sub>3</sub>	<b>181</b> 9-oxo, 12-CHO, Δ <sup>7</sup>	
<b>171</b> 8α, 9β-ΟΗ, 12β -CH <sub>3</sub>	<b>182</b> 9-oxo, 12-OH, Δ <sup>7</sup>	
<b>172</b> 8α, 9α-ΟΗ, 12β -CH <sub>3</sub>	<b>183</b> 9α-OH, 12-COOH, changweikangic acid A	
<b>173</b> 7-oxo, 12 -CH <sub>3</sub> , Δ <sup>8(9)</sup>		

# 3. Concluding remarks

This review provides an extensive list of <sup>13</sup>C NMR spectral data of drimane sesquiterpenes that have been reported by various workers to-date. It has also provided a brief discussion on the substituent effect on the <sup>13</sup>C shielding of the drimane sesquiterpenes. It is evident that direct access to such data will simplify the structure elucidation of new related compounds by data comparison. However, further review of the compounds is still necessary to compile the <sup>1</sup>H NMR spectral data of the compounds.

### **Conflict of interest**

The author declares that there is no conflict of interest.

#### References

Aasen, A.J., Nishida, J., Enzell, C.D., Appel, H.H., 1977. The structure of (11xi, 12xi)-11,12-di (7-drimen11-oxy)-11,12-epoxy-7-drimene. Acta Chem. Scand. 31B, 51-55. Appendino, G., Ozen, H.C., Tagliapietra, S., Cisero, M., 1992. Coumarins from Heptaptera anisoptera. Phytochemistry 31(9), 3211-3213. Aranda, G., Moreno, L., Cortes, M., Prange, T., Maurs, M., Azerad, D., 2001. A new example of  $1\alpha$ -hydroxylation of drimanic terpenes through combined microbial and chemical processes. Tetrahedron 57, 6051-6056.

Ayer, W.A., Craw, P.A., 1989. Metabolites of the fairy ring fungus, Marasmius oreades. Part 2. Norsesquiterpenes, further sesquiterpenes, and agrocybin. Can. J. Chem. 67, 1371-1380. Barrero, A.F., Manzaneda, E.A., Altearejos, J., Salido, S., Ramos, J.M., Simmonds, M.S.J., Blaney, W.M., 1995. Synthesis of biologically active drimanes and homodrimanes from (-)-sclareol. Tetrahedron 51, 7435-7450.

Barrero, F., Cortés, M., Manzaneda, E.A., Cabrera, E., Chahboun, R., Lara, M., Rivas, A.R., 1999. Synthesis of 11,12-epoxydrim-8,12-en-11-ol, 11,12-diacetoxydrimane, and warburganal from (–) -sclareol. J. Nat. Prod. 62, 1488-1491. Bastos, J.K., Kaplan, M.A.C., Gottlieb, O.R., 1999. Drimane-type

sesquiterpenoids as chemosystematic markers of Canellaceae. J. Braz. Chem. Soc. 10, 136-139. Benites, J., Preite, M.D., Cortes, M., 2001. Conversion of

(+)-confertifolin into 11,12-bisnordriman-9-one and (+)- $8\alpha$ H, $9\alpha$ H-11,12-dacetoxydriname. Synth. Commun. 31, 1347-1354.

Butler, M.S., Capon, R.J., 1993. Beyond polygodial: New drimane sesquiterpenes from a Southern Australian marine sponge, Dysidea sp. Aust. J. Chem. 46, 1255-1267.



Chaudhary, S., Thomas, V., Todaro, L., Le Gendre, O., Pecic, S., Harding W.W., 2008. New drimane sesquiterpenoids from Tidestromiao blongifolia. Nat. Prod. Commun. 3, 1747-1750. Chen, Z., Dong, Z., Wen, J., Feng, T., Liu J.A., 2016. New sesquiterpene from the endophytic fungus Nigrospora sphaerica. Rec. Nat. Prod. 10, 307-310.

Claudino, V.D., da Silva, K.C., Filho, V.C., Yunes, R.A., Monache, F.D., Giménez, A., Salamanca, E., Gutierrez-Yapu, D., Malheiros, A., 2013. Drimanes from Drimys brasiliensis with leishmanicidal and antimalarial activity. Mem. Inst. Oswaldo Cruz 108(2), 140-144.

Cuellar, M., Moreno, L., Preite, M., 2003. Regioselective oxidative fragmentation of drimanic terpene alcohols: a short, easy and efficient access to natural and synthetic 11-nordrimane terpene derivatives. ARKIVOC 10, 169-177.

Cunha, F., Fröde, T., Mendes, G., Malheiros, A., Filho, V.C., Yunes, R.A., Calixto, J.B., 2001. Additional evidence for the anti-inflammatory and anti-allergic properties of the sesquiterpene polygodial. Life Sci. 70, 159-169.

Dacunto, M., 2012. Total Synthesis of Terpenoidic Unsatured Dialdehydes and Evaluation of their activity towards TRP Receptors. Ph.D. Thesis, Università Degli Studi di Salerno, Salerno, Italy.

De Bernardi, M., Mellerio, G., Vidari, G., Vita-Finzi, P., Fronza, G., 1980. Fungal metabolites. Part 5. Uvidins, new drimane sesquiterpenes from Lactarius uvidus Fries. J. Chem. Soc. Perkin Transactions. 1, 221-226.

Delgado, V., Armstrong, V., Cortésa, M., Barrero, A.F., 2008. Synthesis of racemic and chiral albicanol, albicanyl acetate and cyclozonarone: Cytotoxic activity of ent-cyclozonarone. Braz. Chem. Soc. 19, 1258-1263.

Derita, M., Montenegro, I., Garibotto, F., Enriz, R., Cuellar, M., Zacchino, S., 2013. Structural requirements for the antifungal activities of natural drimane sesquiterpenes and analogues, supported by conformational and electronic studies. Molecules 18, 2029-2051.

Ding, J.H., Ding, Z.G., Chunyu, W.X., Zhao, J.Y., Wang, H.B., Liu, S.W., Wang, F., 2016. Three new drimane sesquiterpenoids from cultures of the fungus Penicillium sp. J. Asian Nat. Prod. Res. 19, 780-785.

Echeverri, F., Luis, J.G., Torres, F., Quinones, W., Alzate, F., Cardona, G., Archbold, R., Roldan, J., Lahlou, E.H., 1997. Danilol, a new drimane sesquiterpene from Polygonum punctatum leaves. Nat. Prod. Lett. 10, 295-301.

Felix, S., Sandjo, L.P., Opatz, T., Erel, G., 2014. Anti-inflammatory drimane sesquiterpene lactones from an Aspergillus species. Bioorg. Med. Chem. 22, 2912-291.

Fukuyama, Y., Sato, T., Miura, I., Asakawa, Y., 1985. Drimane-type sesqui- and norsesquirterpenoids from Polygonum hydropiper. Phytochemistry 24, 1521-1524.

Gaspar, H., Cutignano, A., Ferreira, T., Calado, G., Cimino, G., Fontana, A., 2008. Biosynthetic evidence supporting the generation of terpene chemodiversity in marine mollusks of the genus Doriopsilla. J. Nat. Prod. 71(12), 2053-2056.

Gören, N., Ulubelen, A., 1988. A sesquiterpene-coumarin ether and an acetylenic compound from Tanacetum heterotomum. Phytochemistry 27(5), 1527-1529.

Grabley, S., Thiericke, R., Zerlin, M., Goert, A., Phillips, S., Zeeck, A., 1996. New albrassitriols from Aspergillus sp. (FH-A 6357). J. Antibiot. 49, 593-595.

Grace, M.H., Lategan, C.A., Mbeunkui, F., Graziose, R.T., Smith, P.J., Raskin, I., Lila, M.A., 2010. Antiplasmodial and cytotoxic activities of drimane sesquiterpenes from Canella winterana. Nat. Prod. Commun. 5(12), 1869-1872. Harinantenaina, L., Matsunami, K., Otsuka, H., Kawahata, M., Yamaguchi, K., Asakawa, Y., 2008. Secondary metabolities of Cinnamosma madagascariensis and their  $\alpha$ -glucosidase

inhibitory properties. J. Nat. Prod. 71, 123-126.

Hashimoto, T., Tori, M., Asakawa, Y., 1989. Drimane-type sesquiterpenoids from the liverwort Makinoa crispate. Phyto-chemistry 28(12), 3377-3381.

He, D., Slebodnick, C., Rakotondraibe, L.H., 2017. Bioactive drimane sesquiterpenoids and aromatic glycosides from Cinnamosma fragrans. Bioorg. Med. Chem. Lett. 27(8), 1754-1759.

He, J.B., Feng, T., Xhang, S., Dong, Z.J., Li, Z.H., Zhu, H.J., Liu, J.K., 2014. Seven new drimane-type sesquiterpenoids from cultures of fungus Phellius tuberculosis. Nat. Prod. Bioprospect. 4, 21-25.

He, J.B., Tao, J., Miao, X.S., Bu,W., Zang, S., Dong, Z.J., Li, Z.H., Feng, T., 2015. Seven new drimane-type sesquiterpenoids from cultures of fungus Laetiporus sulphureus. Fitoterapia 102, 1-6.

Hlubucel, J.R, Aasen A.J., Almqvist, S.O., Enzell, C.R., 1974. Tobacco Chemistry 25. Two new drimane sesquiterpene alcohols from Greek Nicotiana tabacum L. Acta Chem. Scand. B28, 289.

Inocente, E.A., Shaya, M., Acosta, N., Rakotondraibe, L.H., Piermarini, P.M., 2018. A natural agonist of mosquito TRPA1 from the medicinal plant Cinnamosma fragrans that is toxic, antifeedant, and repellent to the yellow fever mosquito Aedes aegypti. PLoS Negl. Trop. Dis. 12, e0006265.

Iranshahi, M., Barthomeuf, C., Bayet-Robert, M., Chollet, P., Davoodi, D., Piacente, S., Rezaee, R., Sahebkar, A., 2014. Drimane-type sesquiterpene coumarins from Ferula gummosa fruits enhance doxorubicin uptake in doxorubicin-resistant human breast cancer cell line. J. Tradit. Complement. Med. 4(2), 118-125.

Jacobsson, U., Muddathir A., 1992. Four biologically active sesquiterpenes of the drimane type isolated from Polygonum glabrum. Phytochemistry 31(12), 4207-4211.

Jansen, B.J.M., 1993. Total synthesis of insect antifeedant drimane sesquiterpenes, Ph.D. Thesis, Agricultural University, Wageningen.

Jansen, B.J.M., de Groot, A., 2004. Occurrence, biological activity and synthesis of drimane sesquiterpenoids. Nat. Prod. Rep. 21, 449-477.

Jeruto, P., Arama, P., Anyango, B., Nyunja, R., Taracha, C., Opiyo S., 2017. Morphometric study of Senna didymobotrya (Fresen.) H. S. Irwin and Barneby in Kenya. J. Nat. Sci. Res. 7(6), 54-69.

Justicia, J., Oltra, J.E., Barrero, A.F., Guadano, A., Gonzalez-Coloma A., Cuerva, J.M., 2005. Total synthesis of 3-hydroxydrimanes mediated by titanocene (III)- evaluation of their antifeedant activity. Eur. J. Org. Chem. 2005, 712-718. Kinoshita, M., Ohtsuka, M., Nakamura, D., Akita, H., 2002. First synthesis of (+)- $\alpha$  - and (+)- $\gamma$ -polypodatetraenes. Chem. Pharm. Bull. 50, 930-934.

Kioy, D., Gray, A.I., Waterman, P.G., 1989. Further drimane sesquiterpenes from the stem bark of Canella winterana. J. Prod. Nat. 52, 174-177.

Kioy, D., Gray, A.I., Waterman, P.G., 1990. A comparative study of the stem-bark drimane sesquiterpenes and leaf volatile oils of Warburgia ugandensis and W. stuhlmannii. Phytochemistry 29, 3535-3538.

Kisiel, W., Stojakowska, A., 2002. A sesquiterpene coumarin ether from transformed roots of Tanacetum parthenium. Phytochemistry 46(3), 515-516.

Kuang, C., Jing, S.X., Liu, Y., Luo, S.H., Li, S.H., 2016. Drimane sesquiterpenoids and isochromone derivative from the endophytic fungi Pestalotiopsis sp. M-23. Nat. Prod. Bioprospect. 6, 155-160.

Kubo, I., Matsumoto, T., Kakooko, A.B., Mubiru, N.K., 1983. Structure of mukaadial, a molluscicide from Warburgia plants. Chem. Lett. 7, 979-980.

Kubo, I., 1995. Antifungal sesquiterpene dialdehydes from the



Warburgia plants and their synergists. In: Atta-ur-Rahman, Ed. Studies in Natural Products Chemistry: Structure and Chemistry (part D). Amsterdam: Elsevier Science, pp. 233-249. Kubo, I., Nakanishi, K., 1977. Insect Antifeedants and Repellents from African Plants. In: Host Plant Resistance to Pest. Hedin P.A., Ed., ACS Symposium Series 62. American Chemical Society, Washington DC. pp. 165-178.

Kuchkova, K.I., Aryku A.N., Dragalin I.P., Vlad P.F., 2005. Synthesis of drim-9 (11)-en- $8\alpha$ -and- $8\beta$ -ols from drimenol. Chem. Nat. Compd. 41, 190-193.

Lagnel, B.M.F., Morin, C., De Groot, A., 2000. Synthesis of drimanes from (+)-larixol. Synthesis 13, 1907-1916.

Lee, C.L., Chiang, L.C., Cheng, L.H., Liaw, C.C., El-Razek, M.H.A., Chang, F.R., Wu, Y.C., 2009. Influenza A (H1N1) antiviral and cytotoxic agents from Ferula assa-foetida. J. Nat. Prod. 72(9), 1568-1572.

Liu, M.S., Liu, C., Zhang, X.P., Sheng, L., Zhang, J.Q., Kang, S.L., 2010. Two new drimane sesquiterpenoids from compound changweikang and their inhibitory activity against nitric oxide production. Chem. Pharm. Bull. 58, 1224-1226.

Lu, Z., Wang, Y., Miao, C., Liu, P., Hong, K., Zhu, W., 2009. Sesquiterpenoids and benzofuranoids from the marine-derived fungus Aspergillus ustus 094102. J. Nat. Prod. 72, 1761-1767. Ludwiczuka, A., Gradsteinb, S.B., Nagashima, F., Asakawa, Y., 2011. Chemosystematics of Porella (Marchantiophyta, Porellaceae). Nat. Prod. Commun. 6(3), 315-321.

Madikane, V.E., Bhakta, S., Russell, A.J., Campbell, W.E., Claridge, T.D.W., Elisha, G., Davies, S.G.D., Smith, P., Sim, E., 2007. Inhibition of mycobacterial arylamine N-acetyltransferase contributes to anti-mycobacterial activity of Warburgia salutaris. Bioorg. Med. Chem.15, 3579-3586.

Mahato, S.B., Kundu, A.P. 1994. 13C NMR spectra of pentacyclic triterpenoids - a compilation and some salient features. Phytochemistry 37, 1517-1575.

Mahmoud, I.I., Kinghorn, A.D., Cordell, G.A., Farnsworth, N.R., 1980. Potential anticancer agents XVI. Isolation of bicyclofarnesane sesquiterpenoids from Capsicodndron dinisii. J. Nat. Prod. 43, 365-371.

Mashimbye, M.J., Maumela, M.C., Drewes, S.E., 1999. A drimane sesquiterpenoid lactone from Warburgia salutaris. Phytochemistry 51, 435-438.

Mathie, K., Lainer, J., Spreng, S., Dawid, C., Andersson, D.A., Bevan, S., Hofmann, T., 2017. Structure-pungency relationships and TRP channel activation of drimane sesquiterpenes in Tasmanian pepper (Tasmannia lanceolata). J. Agric. Food Chem. 65(28), 5700-5712.

Meng, J., Li, Y.Y., Ou, Y.X., Song, L.F., Lu, C.H., Shen Y.M., 2011. New sesquiterpenes from Marasmius cladophyllus. Mycology 2, 30-36.

Mohammadhosseini, M., 2017. The ethnobotanical, phytochemical and pharmacological properties and medicinal applications of essential oils and extracts of different Ziziphora species. Ind. Crops Prod. 105, 164-192.

Mohammadhosseini, M., Sarker, S.D., Akbarzadeh, A., 2017. Chemical composition of the essential oils and extracts of Achillea species and their biological activities: A review. J. Ethnopharmacol. 199, 257-315.

Mohammadhosseini, M., Venditti, A., Sarker, S.D., Nahar, L., Akbarzadeh, A., 2019. The genus Ferula: Ethnobotany, phytochemistry and bioactivities - A review. Ind. Crops Prod. 129, 350-394.

Montagnac, A., Martin, M.T., Debitus, C., Païs, M., 1996. Drimane sesquiterpenes from the sponge Dysìdea fusca. J. Nat. Prod. 59, 866-868.

Montenegro, I., Corral, S.D., Napal, G.N.D., Carpinella, M.C., Mellado, M., Madrid, A.M., Villena, J., Palaciosa, S.M., Cuellar M.A., 2018a. Antifeedant effect of polygodial and drimenol derivatives against Spodoptera frugiperda and Epilachna paenulata and quantitative structure-activity. Pest Manag. Sci. 74, 1623-1629.

Montenegro, I., Madrid, A., Cuellar, M., Seeger, M., Alfaro, J.F., Besoain, X., Martínez J.P., Ingrid Ramirez, I., Olguín, Y., Miryam Valenzuela, M., 2018b. Biopesticide activity from drimanic compounds to control tomato pathogens. Molecules 23, 2053, doi:10.3390/molecules23082053

Montenegro, I., Pino, L., Werner, E., Madrid, A., Espinoza, L., Moreno, L., Villena, J., Cuellar, M., 2013. Comparative study on the larvicidal activity of drimane sesquiterpenes and nordrimane compounds against Drosophila melanogaster til-til. Molecules 18, 4192-4208.

Montenegro, I., Tomasoni, G., Bosio, C., Quiñones, N., Madrid, A., Carrasco, H., Olea, A., Martinez, R., Cuellar, M., Villena, J., 2014. Study on the cytotoxic activity of drimane sesquiterpenes and nordrimane compounds against cancer cell lines. Molecules 19, 18993-19006.

Muñoz, O., Gutierrez, M., Gonzalez, R., Hammann, S., Vetter, W., 2015. Antifungal and insecticidal properties of the phytoconstituents of Drimys winteri (Winteraceae) growing in Chiloe Island (Chile). Nat. Prod. Chem. Res. 3, 4-9.

Nakanishi, K., Kubo, I., 1978. Studies on warburganal, muzigadal and related compounds. J. Chem. 16, 28-31.

Nyaba, Z.N., Murambiwa, P., Opoku, A.R., Mukaratirwa, S., Shode, F.O., Simelane, M.B.C., 2018. Isolation, characterization, and biological evaluation of a potent anti-malarial drimane sesquiterpene from Warburgia salutaris stem bark. Malar. J. 17, 296-303.

Ochieng, C.O., Ishola, I., Opiyo, S.A., Manguro, L.O.A., Owuor, P.O., Wong, K.C. 2013. Phytoecdysteroids from the stem bark of Vitex doniana and their anti-inflammatory effects. Plant Medica. 79, 52-59.

Ochieng, C.O., Opiyo, S.A., Mureka, E.W., Ishola, I., 2017. Cyclooxygenase inhibitory compounds from Gymnosporia heterophylla aerial parts. Fitoterapia 119, 168-174.

Ochung', A.A., Manguro, L.O.A., Owuor, P.O., Jondiko, J.I.O., Nyunja, R.A., Akala, H., Mwinzi, P., Opiyo, S.A., 2015. Bioactive carbazole alkaloids from Alysicarpus Ovalifolius (Schumach). J. Korean. Soc. Appl. Biol. Chem., 58(6), 839-846.

Ochung', A.A., Owuor, P.O., Manguro, L.O.A., Ishola, I.O., Nyunja, R.A., Ochieng, C.O., Opiyo, S.A., 2018. Analgesics from Lonchocarpus eriocalyx Harms. Trends Phytochem. Res. 2(4), 253-260.

Odyek, O., Olila, D., Albrecht, C., Dagne, E., 1993. Muzigadial, a cytotoxic sesquiterpine from Warburgia ugandensis. Proceedings of the 5th NAPRECA Symposium on Natural Products, September19 - 23, 1993, Antananarivo Madagascar, p. 160.

Ohloff, G., Giersch, W., 1985. Structure-activity relationships in odor perception of drimane derivatives, CCACAA, 58, 491-509.

Opiyo, S.A., Manguro, L.A.O., Owuor, P.O., Ateka, E.M., 2017. Triterpenes from Elaeodendron schweinfurthianum and their antimicrobial activities against crop pathogens. Am. J. Chem. 7(3), 97-104.

Opiyo, S.A., 2011. Development of a Multiplex PCR Technique for Simultaneous Detection of Sweet Potato Viruses and Evaluation of Traditional Medicinal Plant Extracts for Antimicrobial Activity Against the Crop Pathogens. Ph.D. Thesis, Maseno University, Kenya.

Opiyo, S.A., Manguro, L.O.A., Okoth, D.A., Ochung, A.A., Ochieng, C.O., 2015. Biopesticidal extractives and compounds from Warburgia ugandensis against maize weevil (Sitophilus zeamais). Nat. Prod. J. 5, 236-243.

Opiyo, S.A., Manguro, L.O.A., Owuor P.O., Ateka, E.M., Lemmen, P., 2011.  $7\alpha$ -Acetylugandensolide and antimicrobial properties of Warburgia ugandensis extracts and isolates against sweet potato pathogens. Phytochem Lett. 4, 161-165.



Panasenko A., Gorincioi E.C., Aricu A.N., Barcari E.A., Deleanu K., Vlad PF., 2004. 1H and 13C NMR spectra of some drimanic sesquiterpenoids. Russ. Chem. Bull. Int. Ed. 53, 2700-2705. Paul, V.J., Seo, Y., Cho, K.W., Rho, J.R., Shin, J., Bergguist, P.R.J., 1997. Sesquiterpenoids of the drimane class from a sponge of the genus Dysidea. J. Nat. Prod. 60, 1115-1120. Pittayakhajonwut, P., Dramae, A., Intaraudom, C., Boonyuen, N., Nithithanasilp, S., Rachtawee, P., Laksanacharoen, P., 2011. Two new drimane sesquiterpenes, fudecadiones A and B, from the soil fungus Penicillium sp. BCC 17468. Planta Med. 77, 74-76.

Rajab, M.S., Ndegwa, J.S., 2000.  $11\alpha$ -Hydroxy muzigadiolide, a novel drimane sesquiterpenes from the stem bark of Warburgia ugandensis. Ethiopia Chem. Soc. Bull. 14, 45-49. Ramirez, H.E., Cortes, M.M., Agosin, E., 1993. Bioconversion of drimenol into 3 $\beta$ -hydroxydrimanes by Aspergillus niger. effect of culture additives. J. Nat. Prod. 56, 762-764. Reddy, P.P., Tiwari, A.K., Rao, R.R., Madhusudhana, K., Rao, V.R.S., Ali, A.Z., Babu, K.S., Rao, J.M., 2009. New labdane diterpenes as intestinal  $\alpha$ -glucosidase inhibitor from antihyperglycemic extract of Hedychium spicatum (Ham. Ex Smith) rhizomes. Bioorg. Med. Chem. Lett. 19(9), 2562-2565. Rodríguez, B., Zapata, N., Medina, P., Viñuela, E., 2005. A complete 1H and 13C-NMR data assignment for four drimane sesquiterpenoids isolated from Drimys winteri. Magn. Reson. Chem. 43, 82-84.

Rukachaisirikul, V., Khamthong, N., Sukpondma, Y., Phongpaichit, S., Hutadilok-Towatana, N., Graidist, P., Sakayaroj, J., Kirtikara, K., 2010. Cyclohexene, diketopiperazine, lactone and phenol derivatives from the sea fan-derived fungi Nigrospora sp. PSU-F11 and PSU-F12. Arch. Pharm. Res. 33, 375-380. Sakio, Y., Hirano, Y.J., Hayashi, M., Komiyama, K., Ishibashi, M., 2001. Dendocarbins A-N, new drimane sequiterpenes from the nudibranch Dendrodoris carbunculosa. J. Nat. Prod. 64, 726-731.

Santoso, B.B., Hernandez, H.P., Rodriguez, E.B., Dalmacio, I.F., 2018. Two antimicrobial compounds drimane sesquiterpene polygodial and 11-hydroxydrim-8-en-7-one from the stem bark of Drimys arfakensis Gibbs. (Winteraceae). Mal. J. Fund. Appl. Sci. 1, 150-154.

Seeram, N.P., Francis, L.S., Needham, O.L., Jacobs, H., McLean, S., Reynolds, W.F., 2003. Drimane and bisabolane sesquiterpenoids from Cinnamodendron corticosum (Canellaceae). Biochem. Syst. Ecol. 31, 637-640.

Shiono, Y., Hiramatsu, F., Murayama, T., Koseki, T., Funakoshi, T., Ueda, K., Yasuda, H., 2007. Two drimane-type sesquiterpenes, strobilactones A and B, from the liquid culture of the edible mushroom strobilurus ohshimae. Z. Naturforsch. 62, 1585-1589.

Skiredj, A., Beniddir, M.A., Evanno, L., Poupon, E., 2016. Mimicking the main events of the biosynthesis of drimentines: synthesis of  $\Delta 8'$ -isodrimentine A and related compounds. Eur. J. Org. Chem. 2016, 2954-2958.

Sultana, R., Hossain, R., Adhikari, A., Ali, Z., Yousuf, S., Choudhary, M.I., Ali, M.Y., Zaman, M.S., 2011. Drimane-type sesquiterpenes from Polygonum hydropiper. Planta Med. 77, 1848-1851.

Toshima, H., Oikawa, H., Toyomasu, T., Sassa, T., 2001. Total synthesis of (+)-albicanol and (+)-albicanyl acetate. Biosci. Biotechnol. Biochem. 65, 1244-1247.

Tosun, F., Beutler, J.A., Ransom, T.T., Miski, M., 2019. Anatolicin, a highly potent and selective cytotoxic sesquiterpene coumarin from the root extract of Heptaptera anatolica. Molecules 24, 1153-1160.

Toyota, M., Ooiso, Y., Kusuyama, T., Asakawa, Y., 1994. Drimane-type sesquiterpenoids from the liverwort Diplophyllum serrulatum. Phytochemistry 35(5), 1263-1265.

Vieira, D.R., Amaral, F.M., Maciel, M.C, Nascimento, F.R.,

Libério, S.A., Rodrigues, V.P., 2014. Plant species used in dental diseases: ethnopharmacology aspects and antimicrobial activity evaluation. J. Ethnopharmacol. 155(3), 1441-1449. Vlad, P.F., Coltsa, M.N., Aricu, A.N., Ciocarlan, A.G., Gorincioi, E.C., Edu, C.G., Deleanu, C., 2006. Photooxidative dehydrogenation of  $\Delta$ 8-drimen-and  $\Delta$ 8-11-homodrimen-7-ones into  $\alpha, \alpha'$ -dienones. Russ. Chem. Bull. 55, 703-707. Vlad, P.F., Gorincioi, E.C., Coltsa, M.N., Deleanu, C., 2000.

Synthesis of isodrimenin from drim-8-en-7-one. Russ. Chem. Bull. 49(3), 546-548.

Wahlberg, I., Eklund, A.M., Nishida, T., Enzell, C.R., 1981. Tobacco chemistry. Two new nor-drimanes from Greek tobacco. Acta Chem. Scand. B. 35, 307-310.

Wansi, J.D., Sewald, N., Nahar, L., Martin, C., Sarker, S.D., 2019. Bioactive essential oils from the Cameroonian rain forest: A review - Part II. Trends Phytochem. Res. 3(1), 3-52.

Wansi, J.D., Sewald, N., Nahar, L., Martin, C., Sarker, S.D., 2018. Bioactive essential oils from the Cameroonian rain forest: A review - Part I. Trends Phytochem. Res. 2(4), 187-234. Wayman, K.A., de Lange, P.J., Larsen. L., Sansom, C.E., Perry, N.B., 2010. Chemotaxonomy of Pseudowintera: sesquiterpene dialdehyde variants are species markers. Phytochemistry 71(7), 766-72.

World Health Organization, WHO, 2014. Traditional Medicine Strategy 2014-2023, 2014. DOI: http://aps.who.int7iris/bit-stream710665/92455/119789241506090eng.pdf?ua=1.

Wube, A.A., Franz, B., Gibbons, S., Asres, K., 2005. Sesquiterpenes from Warburgia ugandensis and their antimycobacterial activity. Phytochemistry 66, 2309-2315.

Wube, A.A., Bucar, F., Gibbons S, Asres, K., Rattreay, L., Croft, S.L., 2010. Antiprotozoal activity of drimane and coloratane sesquiterpenes towards Trypanosoma brucei rhodesiense and Plasmodium falciparum in vitro. Phytother. Res. 24, 1468-1472.

Xiao, J., Lin, L., Hu, J., Jiao, F., Duan, D., Zhang, Q., Tang, H., Gao, J., Wang, X., 2017. Highly oxygenated caryophyllene-type and drimane-type sesquiterpenes from Pestalotiopsisadusta, an endophytic fungus of Sinopodophyllum hexandrum. RSC Adv. 7, 29071-29079.

Xu, D., Sheng, Y., Zhou, Z.Y., Liu, R., Leng, Y., Liu, J.K., 2009a. Sesquiterpenes from cultures of the basidiomycete Clitocybe conglobata and their 11D-hydroxysteroid dehydrogenase inhibitory activity. Chem. Pharm. Bull., 57, 433-435. Xu, M., Litaudon, M., Krief, S., Martin, M.T., Kasenene, J.,

Kiremire, B., Dumontet, V., Guéritte, F., 2009b. A new drimane-type sesquiterpenoid from Warburgia ugandensis. Molecules 14, 3844-3850.

Yang, X.Y., Feng, T., Ding, J.H., Li, Z.H.,Li, Y., Fan, Q.Y., Liu, J.K., 2013. Two new drimane sesquiterpenoids from cultures of the basidiomycete Trichaptum biforme. Nat. Prod. Bioprospect. 3, 154-157.

Yonemura, Y., Ohyama, T., Hoshino, T., 2012. Chemo-enzymatic syntheses of drimane-type sesquiterpenes and the fundamental core of hongoquercin meroterpenoid by recombinant squalene-hopene cyclase. Org. Biomol. Chem. 10, 440-446.

Zhao, Z.Z., Chen, H.P., Feng, T., Li, Z.H., Liu, J.K., 2014. Four new sesquiterpenoids from cultures of the fungus Phellinidium sulphurascens. Nat. Prod. Bioprospect. 5, 23-28.