



ORIGINAL ARTICLE

Open Access



Leveraging off higher plant phylogenetic insights for antiparasmodial drug discovery

Phanankosi Moyo¹ , Luke Invernizzi¹ , Sephora M. Mianda¹ , Wiehan Rudolph¹ , Warren A. Andayi² , Mingxun Wang³ , Neil R. Crouch^{4,5} and Vinesh J. Maharaj^{1*}

Abstract

The antimalarial drug-resistance conundrum which threatens to reverse the great strides taken to curb the malaria scourge warrants an urgent need to find novel chemical scaffolds to serve as templates for the development of new antimalarial drugs. Plants represent a viable alternative source for the discovery of unique potential antiparasmodial chemical scaffolds. To expedite the discovery of new antiparasmodial compounds from plants, the aim of this study was to use phylogenetic analysis to identify higher plant orders and families that can be rationally prioritised for antimalarial drug discovery. We queried the PubMed database for publications documenting antiparasmodial properties of natural compounds isolated from higher plants. Thereafter, we manually collated compounds reported along with plant species of origin and relevant pharmacological data. We systematically assigned antiparasmodial-associated plant species into recognised families and orders, and then computed the resistance index, selectivity index and physicochemical properties of the compounds from each taxonomic group. Correlating the generated phylogenetic trees and the biological data of each clade allowed for the identification of 3 'hot' plant orders and families. The top 3 ranked plant orders were the (i) Caryophyllales, (ii) Buxales, and (iii) Chloranthales. The top 3 ranked plant families were the (i) Ancistrocladaceae, (ii) Simaroubaceae, and (iii) Buxaceae. The highly active natural compounds ($IC_{50} \leq 1 \mu M$) isolated from these plant orders and families are structurally unique to the 'legacy' antimalarial drugs. Our study was able to identify the most prolific taxa at order and family rank that we propose be prioritised in the search for potent, safe and drug-like antimalarial molecules.

Keywords Natural products, Plants, Phylogenetics, Malaria, Drug-resistance, 'Hot' plants

1 Introduction

Malaria is a vector-borne tropical disease caused by unicellular protozoan parasites of the genus *Plasmodium* [1]. Despite conspicuous progress in controlling and managing malaria, the disease remains a serious public health challenge. Malaria is currently endemic in 84 countries, with the World Health Organisation (WHO) reporting the African Region as the most afflicted by this disease [2]. In 2021 there was a total of 247 million clinical malaria cases and 619,000 malaria-induced fatalities globally, with the African Region accounting for 95% of the reported cases and fatalities. Current antimalarial drugs are becoming less effective due to the emergence and spread of drug-refractory *Plasmodium* parasite strains [3]. These strains arise as a product of mutations,

*Correspondence:

Vinesh J. Maharaj
vinesh.maharaj@up.ac.za

¹ Biodiscovery Center, Department of Chemistry, Faculty of Natural and Agricultural Sciences, University of Pretoria, Private Bag X 20, Hatfield, Pretoria 0028, South Africa

² Department of Physical and Biological Sciences, Murang'a University of Technology, Murang'a, Kenya

³ Computer Science and Engineering, University of California Riverside, 900 University Ave, Riverside, CA 92521, USA

⁴ Biodiversity Research and Monitoring Directorate, South African National Biodiversity Institute, Berea Road, P.O. Box 52099, Durban 4007, South Africa

⁵ School of Chemistry and Physics, University of KwaZulu-Natal, Durban 4041, South Africa



Springer

© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

most notably due to either DNA replication errors or damage induced by reactive oxygen species [4, 5]. These mutations give rise to phenotypes with changes in, for example, either drug targets or transporters which confer *Plasmodium* parasites with resistance to antimalarial drugs. This resistance phenomenon has been observed for all previously used antimalarials [6], including the current WHO-recommended first-line treatment drugs for malaria, namely artemisinin-based combination therapy (ACT) [6]. This highlights the need to discover and develop new and alternative malaria treatment regimens.

One promising strategy for discovering new curative antimalarial compounds is the identification of plants that produce compounds with antiplasmodial activity. Plants have evolved to produce a diverse array of chemical compounds to defend themselves against pathogens and parasites, many of which have potential medicinal properties. This has made them a reliable source for the discovery of privileged chemical scaffolds, which have served as a foundation for developing a plethora of pharmaceutical agents [7]. Some of the linchpin malaria chemotherapeutics were likewise discovered from plants. From a *Cinchona* species (Rubiaceae), the alkaloid quinine was isolated. This compound served as a template from which derivatives, including chloroquine, were synthesised. Lapachol, a naphthoquinone first isolated in 1882 from the bark of *Tabebuia avellanedae* (Bignoniaceae), served as a scaffold which inspired the development of the antimalarial drug atovaquone. Similarly, from the Chinese herb *Artemisia annua* (Asteraceae), the sesquiterpene lactone artemisinin was isolated and semi-synthesised to form prolific fast-acting derivatives, namely artemether, dihydroartemisinin and artesunate which are the core constituents of the ACT regimen [8]. Over the last century, these plant-derived antimalarials have saved millions of lives [9–11]. In view of this, there is merit in the continued investigation of plants in search of novel antimalarial agents to redress the drug-resistance scourge.

Given the diversity and expansiveness of the plant kingdom (*ca.* 370,000 flowering plant species [12]) and limited research resources, there is a need for a rational strategy to streamline and focus drug screening projects on selected plant species. Adoption of such strategies is envisaged to expedite antimalarial drug discovery by simplifying the plant screening process. Moreover, this is anticipated to come with the added advantage of an increased likelihood of discovering promising leads (hits) as research resources will be focused on the ‘hot’ taxonomic groups (i.e., taxa with an overrepresentation of active compounds) [13, 14]. One logical approach adopted in some drug discovery projects that prioritise plant subjects for evaluation, is phylogenetic analysis

[15–17]. Such phylogenetic analyses allow for identification of ‘hot’ plant genera, families or orders that have been demonstrated to produce bioactive compounds against specific therapeutic targets [18]. This concept emanates from the premise that phylogeny and biosynthetic pathways are correlated; therefore, the production of specific bioactive natural products with peculiar biological properties will likely be common to closely related plant species at the level of genus, family, or families within an order [14]. In line with this principle various members of the filamentous bacterial genus *Streptomyces* have yielded an array of secondary metabolites (including tetracyclines, aminoglycosides and macrolides) with commercially useful antimicrobial activity [19, 20]. Similarly, amongst higher plants, the Amaryllidaceae plant family is well established to exclusively produce specific alkaloids, including the lycorine-type alkaloids, which exhibit a distinct pharmacological profile [21]. The correlation of phylogenetic analysis and pharmacological data allows us to effectively identify ‘hot’ plant taxonomic groups of specific interest to man based on what is experimentally known about compounds isolated from plant species in those plant orders and families. From these taxonomic groups, closely related, untapped species can be rationally prioritised for pharmacological evaluation [18].

Over recent decades, numerous natural compounds isolated from different plant species have been evaluated for their *in vitro* antiplasmodial activity. This study aimed to use phylogenetic analysis to identify ‘hot’ plant orders, and families that: (i) produce active antiplasmodial compounds ($IC_{50} \leq 10 \mu M$) with (ii) acceptable resistance index ($RI \leq 10$), (iii) a selectivity index ($SI \geq 10$), and (iv) drug-like properties. We collated data on active and inactive antiplasmodial plant-derived compounds from literature published between 1964 and 2021. We determined, including through resolution of synonymy, the identity of plant species yielding antiplasmodial isolates and used the information to construct phylogenetic trees, which were correlated to quantified antiplasmodial and cytotoxicity data. From the generated trees, we were able to establish the distribution patterns of plant species in different ‘hot’ plant families and orders. We believe this analysis will facilitate the selection of taxa warranting further evaluation in antimalarial drug discovery programs, to optimize project outcomes.

2 Results, discussion and conclusion

2.1 Descriptive analysis: articles evaluated, plant taxonomy, and antiplasmodial screening

A PubMed database query using the key phrase “*Plasmodium falciparum* and natural product”, limiting the search to the abstract, yielded a total of 3863 articles (Fig. 1).

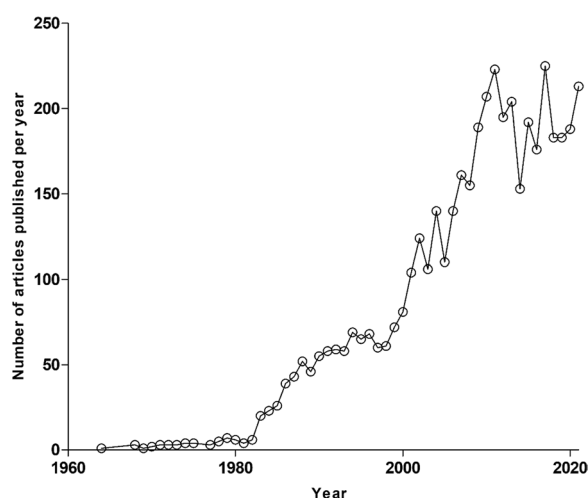


Fig. 1 Number of articles published per year (1964 to 2021) on “*Plasmodium falciparum* and natural product”. The annual number of publications on the topic of “*Plasmodium falciparum* and natural product” has evidently increased over the years within the last decade, being the most productive in that regard. These search results were realised following the query made on PubMed and limited to abstracts

These articles were published within a 58-year period ranging from 1964 to 2021 (Fig. 1). From this pool of articles, duplicates and publications reporting on non-higher plant-derived natural products were excluded. Furthermore, studies where natural compounds were neither isolated nor screened in vitro against *P. falciparum*, were excluded. Following the application of these exclusion criteria, we filtered to a total subset of 455 articles.

From the relevant 455 articles, 2426 natural compounds, reportedly either active or inactive in vitro against *P. falciparum* parasites were manually compiled. These compounds were collected along with components of plant species from which they were initially isolated, and their respective antiparasitic activities were determined. These molecules were isolated from 439 plant species belonging to 99 vascular plant families referred to 37 plant orders (Table 1). This total number of plant species only represents a small fraction (*ca.* 0.1%) of all known higher plant species worldwide. The *ca.* 2400 plant-derived compounds which our study was limited to is substantially less than that analysed in a similar study by Zhu et al. [15] where *ca.* 31,000 compounds were considered. In their study, Zhu and colleagues examined the species-distribution of 939 clinically approved natural product-derived drugs, 369 clinical candidates, and 119 preclinical candidates. Furthermore, 13,548 marine derived natural products and 19,721 bioactive secondary metabolites were included in their study [15]. The natural product derived drugs and bioactive compounds

considered were from several sources including plants, microorganisms, and marine organisms [15]. Furthermore, these were drugs and bioactive compounds for several therapeutic areas [15]. Disease-area focused investigations have previously analysed the number of compounds consistent with our study, although not for antiparasitic activity. For example, in their investigation to examine the phylogenetic distribution of anticancer drugs, Li et al. [22] analysed 207 natural product derived drugs. From a malaria drug discovery standpoint our study is, to the best of our knowledge, the first to extensively examine the relationship between phylogeny and antiparasitic activity of natural product compounds. Earlier approaches have focused on compound classes in relation to antiparasitic activity, exemplified by the study of Egieyeh et al. [23] which used cheminformatics profiling to prioritise natural products for antimalarial drug discovery. In their study they managed to analyse 1040 natural product compounds isolated from plants, microorganisms, and marine sources [23].

Of the 2400 compounds analysed in our study, the Asterids and Rosid lineages were the most overrepresented clades with 6 and 10 plant orders, respectively (see Table 3). The plant order with the greatest number of different accepted plant species investigated was the Sapindales ($n=65$), closely followed by Asterales ($n=48$) and Gentianales ($n=55$). The Chloranthales (4%), Cupressales (1.8%), Canellales (1.75%), Buxales (1.54%), and Sapindales (0.95%) were the most investigated relative to the total number of accepted plant species known to occur in those orders. In contrast, the Asparagales (0.02%), and Alismatales (0.02%) were evidently the orders least investigated, considering their species richness (Table 1).

Compounds isolated from plant species in the different taxonomic groups were primarily assessed for activity against the 3D7 ($n=426$), D6 ($n=254$), and NF54 ($n=182$) intra-erythrocytic asexual *P. falciparum* parasite drug-sensitive (D-S) strains. In vitro evaluation of potency against the intra-erythrocytic asexual *P. falciparum* parasite drug-resistant (D-R) parasites was predominantly carried out on the K1 ($n=689$), W2 ($n=399$) and Dd2 ($n=317$) strains (Table 2).

2.2 Phylogenetic analyses correlated to biological data

Following identification to species rank of taxa yielding compounds tested for antiparasitic activity phylogenetic trees (cladograms) of higher plant orders and plant families were constructed using the NCBI Taxonomy database [24] and graphically displayed using an online tool, viz., the ‘interactive Tree of Life’ (iTOL) (Figs. 2 and 3) [25]. The constructed trees are consistent with the taxonomic classification and nomenclature reflected in ‘The World Flora Online’ [26]. The relationship between

Table 1 Taxonomic representation of plant species yielding compounds subsequently investigated for antiplasmodial activity, arranged by Order

Plant Order	No. of accepted ^a		No. of investigated (ca.) ^b		% Investigated ^c	
	Species	Families	Species	Families	Species	Families
Alismatales	4476	14	1	1	0.02	7.14
Apiales	5935	7	5	1	0.08	14.29
Asparagales	39,041	14	9	3	0.02	21.43
Asterales	37,448	11	48	1	0.13	9.09
Boraginales	3522	1	1	1	0.03	100.00
Brassicales	5172	17	2	3	0.04	17.65
Buxales	130	1	2	1	1.54	100.00
Canellales	114	2	2	2	1.75	100.00
Caryophyllales	12,797	41	13	6	0.10	14.63
Celastrales	1385	2	6	1	0.43	50.00
Chloranthales	73	1	3	1	4.11	100.00
Commelinales	928	5	1	1	0.11	20.00
Cornales	707	7	2	2	0.28	28.57
Cucurbitales	3368	8	1	1	0.03	12.50
Cupressales	222	3	4	2	1.80	66.67
Dioscoreales	896	3	1	1	0.11	33.33
Dipsacales	1348	2	1	1	0.07	50.00
Ericales	15,376	22	6	4	0.04	18.18
Fabales	25,024	4	35	1	0.14	25.00
Fagales	1629	7	6	3	0.37	42.86
Gentianales	23,061	5	55	5	0.24	100.00
Lamiales	28,037	24	27	9	0.10	37.50
Laurales	3831	7	8	3	0.21	42.86
Lycopodiales	425	1	1	1	0.24	100.00
Magnoliales	3228	6	21	3	0.65	50.00
Malpighiales	19,119	36	45	11	0.24	30.56
Malvales	7404	10	5	3	0.07	30.00
Myrtales	14,510	9	17	3	0.12	33.33
Oxalidales	2093	7	3	3	0.14	42.86
Piperales	4276	3	8	2	0.19	66.67
Poales	25,176	14	6	2	0.02	14.29
Proteales	2027	4	2	2	0.10	50.00
Ranunculales	6091	7	13	4	0.21	57.14
Rosales	10,894	9	7	3	0.06	33.33
Santalales	2513	7	2	1	0.08	14.29
Sapindales	6826	9	65	6	0.95	66.67
Zingiberales	2971	8	5	1	0.17	12.50

^a The No. of accepted plant species and families per plant order are as reported in the World Flora Online Plant List database (<https://wfoplantlist.org/plant-list/>), which was also sourced to resolve issues of synonymy and nomenclature at all taxonomic ranks

^b The total no. of investigated plant species and families per plant order are from data collated in this study

^c The % investigated for each order was calculated by dividing the No. of investigated species and families by the No. of accepted species and families, respectively

antiplasmodial activity and phylogenetic relationships at order and associated family levels was then determined and expressed as “hit rates (HR) %” for taxonomic groups with ≥ 10 compounds isolated from them. The HR were calculated by dividing the number of compounds with

an $IC_{50} \leq 10 \mu M$ by the total number of compounds isolated and experimentally evaluated for activity against either the D-S or D-R plasmodia. Plant taxa were correlated to the calculated HR of their compounds. Given the extensive diversity of higher plants globally, we consider

Table 2 Top 5 D-S and D-R intra-erythrocytic asexual *P. falciparum* parasite strains most targeted for in vitro antiparasmodial screening of plant-derived compounds in reports considered

<i>P. falciparum</i> strain (D-S)	No. of compounds screened	<i>P. falciparum</i> strain (D-R)	No. of compounds screened
3D7	403	K1	672
D6	257	W2	372
NF54	183	Dd2	295
D10	98	FCB1	189
FCA	81	FCM29	54

it prudent to strategically focus discovery phase research on either ‘hot’ plant orders, or ‘hot’ families which yield compounds with high HR. This is assumed to increase the likelihood of successfully identifying active antiparasmodial compounds within a short time frame.

Generally, compounds from most plant orders showed high HR of 35% and 43% against asexual D-S and D-R *P. falciparum* parasites, respectively (Fig. 2). Compounds isolated from the Buxales had the highest HR (96%, n (number of compounds)=25). This was closely followed by the Caryophyllales (HR=79%, n =53) and Laurales (HR=67%, n =12) (Fig. 2). The lowest HR were noted for the Dioscoreales (HR=0%, n =14), Ericales (HR=13%, n =23), and Magnoliales (HR=16%, n =91). Against D-R strains, the Celastrales (HR=92%, n =12) and Buxales (HR=91%, n =11) were found to have the highest HR, whereas the Dioscoreales (HR=0%, n =14), and Brassicales (HR=10%, n =10) presented the lowest HR against this *Plasmodium* form. Noteworthy plant orders with markedly different HR in relation to D-S and D-R parasites were the Celastrales (51% difference), Brassicales (30% difference) and Piperales (22% difference).

Compounds isolated from the Simaroubaceae (in Sapindales) demonstrated the highest HR (100%, n =19) against the D-S parasites (Fig. 3). This plant family was closely followed by the Buxaceae (in Buxales) (HR=96%, n =25), Ancistrocladaceae (in Caryophyllales) (HR=76%, n =42), and Lauraceae (in Laurales) (HR=70%, n =10) (Fig. 3). The lowest HR were noted for the Dioscoreaceae (in Dioscoreales) (HR=0%, n =14), Malvaceae (in Malvales) (HR=0%, n =10), and Bignoniaceae (in Lamiales) (HR=7%, n =14). Against D-R strains, the Ancistrocladaceae (HR=96%, n =45) had the highest HR, marginally more than that for the Celastraceae (in Celastrales) (HR=92%, n =12), Buxaceae (HR=91%, n =11) and Simaroubaceae (HR=91%, n =55). The Dioscoreaceae (HR=0%, n =14), Scrophulariaceae (in Lamiales) (HR=8%, n =25), Rubiaceae (in Gentianales) (HR=15%,

n =53), Phyllanthaceae (in Malpighiales) (HR=16%, n =18), and Malvaceae (HR=16%, n =19) demonstrated the lowest HR against the D-R parasites (Fig. 3). Plant families with notably different HR against the D-S and D-R parasites were Celastraceae (51% difference), and Loganiaceae (in Gentianales) (29% difference).

Overall, from this preliminary analysis, the Buxales and Caryophyllales have consistently emerged as the ‘hottest’ plant orders. The Simaroubaceae, Buxaceae, and Ancistrocladaceae have emerged as the ‘hottest’ plant families, routinely displaying high HR against both the D-S and D-R *Plasmodium* parasites (Fig. 4).

2.3 Antiparasmodial activity and cytotoxicity of compounds isolated from different plant orders

To further assess the productivity of the different plant orders and families, we expanded our analysis by determining the % of compounds, in each plant order and family, classified as either highly active (HA) ($IC_{50} \leq 1 \mu M$), moderately active (MA) ($10 \mu M \geq IC_{50} > 1 \mu M$) or poorly active (PA) ($IC_{50} > 10 \mu M$) (Table 3). We anticipate that research that prioritises plant orders or families that produce mainly HA compounds, will yield more rewarding leads. Furthermore, considering the need to address the resistance phenomenon, we examined the resistance index (RI) of compounds as an indicator of their efficacy against the D-R strains relative to D-S *Plasmodium* parasite strains. Additionally, as a proxy indicator for the preference of compounds to compromise *Plasmodium* parasite proliferation ahead of that of mammalian cell lines, we assessed the selectivity index (SI) of the compounds investigated per plant order and family. We consider that plant orders and families producing compounds with an $RI \leq 10$ and an $SI \geq 10$ should be preferentially prioritised for investigation (as per guidelines provided by the Medicines for Malaria Venture (MMV)—<https://www.mmv.org/>).

This analysis showed that many of the compounds isolated from plant species in different plant orders were found to be PA. Exceptions to this were the plant orders Caryophyllales (n =53), with 55% of its compounds found to be HA against *P. falciparum* D-S strains (Table 3). Similarly, against D-R strains, many compounds (45%) from the Caryophyllales (n =62) were classified as HA. Likewise, most of compounds (46 to 75%) from the orders Chloranthales (n =44), Buxales (n =11) and Celastrales (n =12) were classified as being HA against the D-R strains. Despite receiving considerable attention, the majority (ranging from 61 to 80%) of the compounds isolated from orders Asterales (n =151), Gentianales (n =192), Lamiales (n =56), Sapindales (n =168), Malpighiales (n =86), Fabales (n =138), and Magnoliales (n =91) were classified as PA against D-S *P. falciparum*

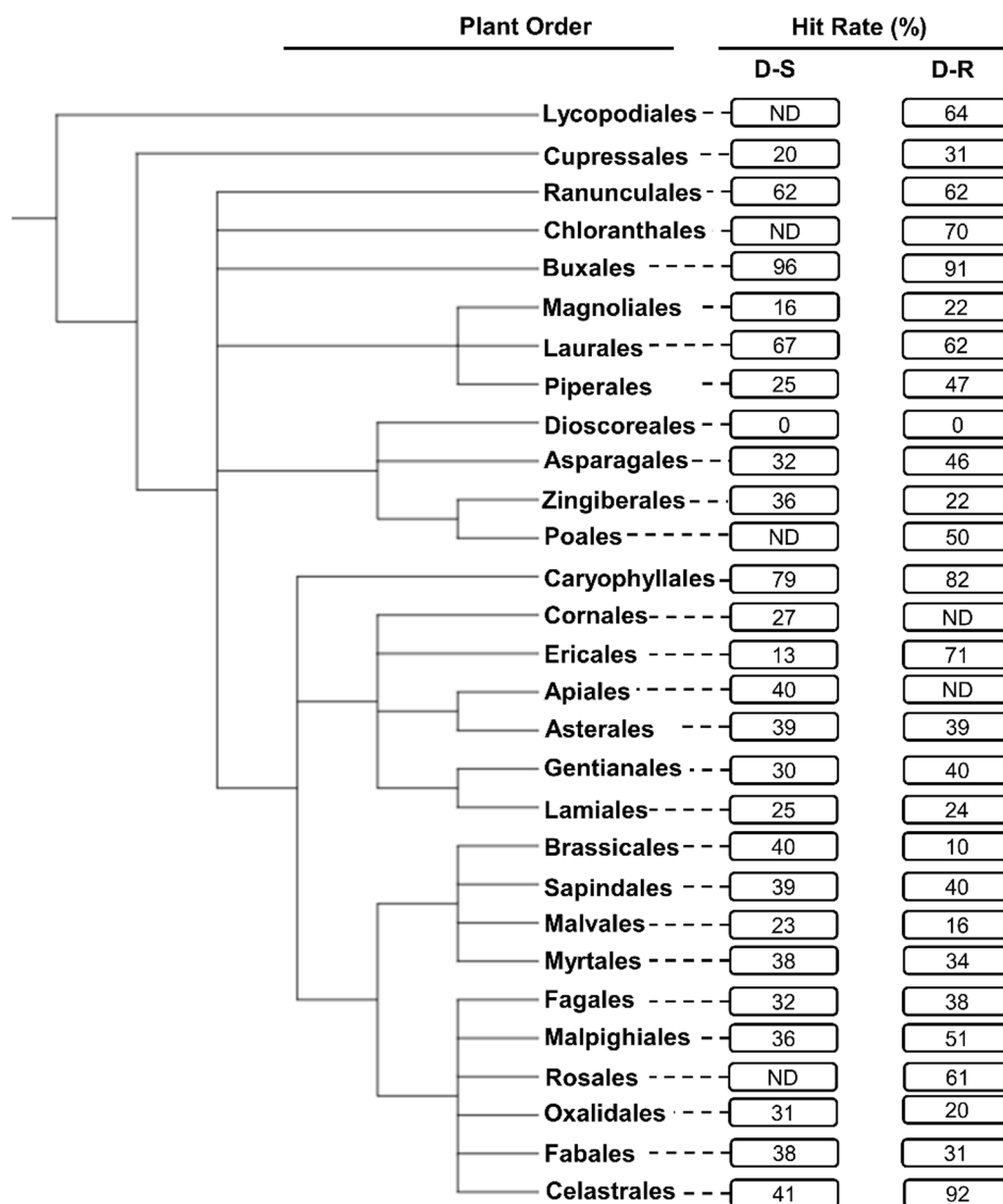


Fig. 2 Phylogenetic tree of plant orders investigated in vitro for their activity against intra-erythrocytic asexual *P. falciparum* parasites. The tree was generated using NCBI Taxonomy and processed using iTOL. ND not determined: This applies for plant orders with < 10 compounds isolated from them and subsequently evaluated for their antiplasmodial activity. The hit rate (HR) is the % of compounds with an $IC_{50} \leq 10 \mu M$ for each plant order. D-S—drug-sensitive. D-R—drug-resistant

strains. This pattern was noted for the same orders, namely Asterales ($n=145$), Gentianales ($n=190$), Lamiales ($n=98$), Sapindales ($n=287$), Malpighiales ($n=186$), Fabales ($n=177$), and Magnoliales ($n=117$), against D-R strains with most of the compounds (ranging from 49 to 78%) being classified as PA. Generally, most of the compounds in all plant orders demonstrated an acceptable RI, i.e., ≤ 10 . In addition to having most of their compounds classified as either HA and MA, remarkably,

many molecules (>70%) from the Buxales ($n=31$), Chloranthales ($n=13$), and Caryophyllales ($n=31$) showed a good SI, i.e., ≥ 10 (Table 3).

Consistent with observations for the plant orders, most compounds isolated from most plant families were classified as PA (Table 4). In variance with this generalisation were the Ancistrocladaceae ($n=42$) and Simaroubaceae ($n=19$), from which most compounds (52 and 74%, respectively) were classified as HA against the

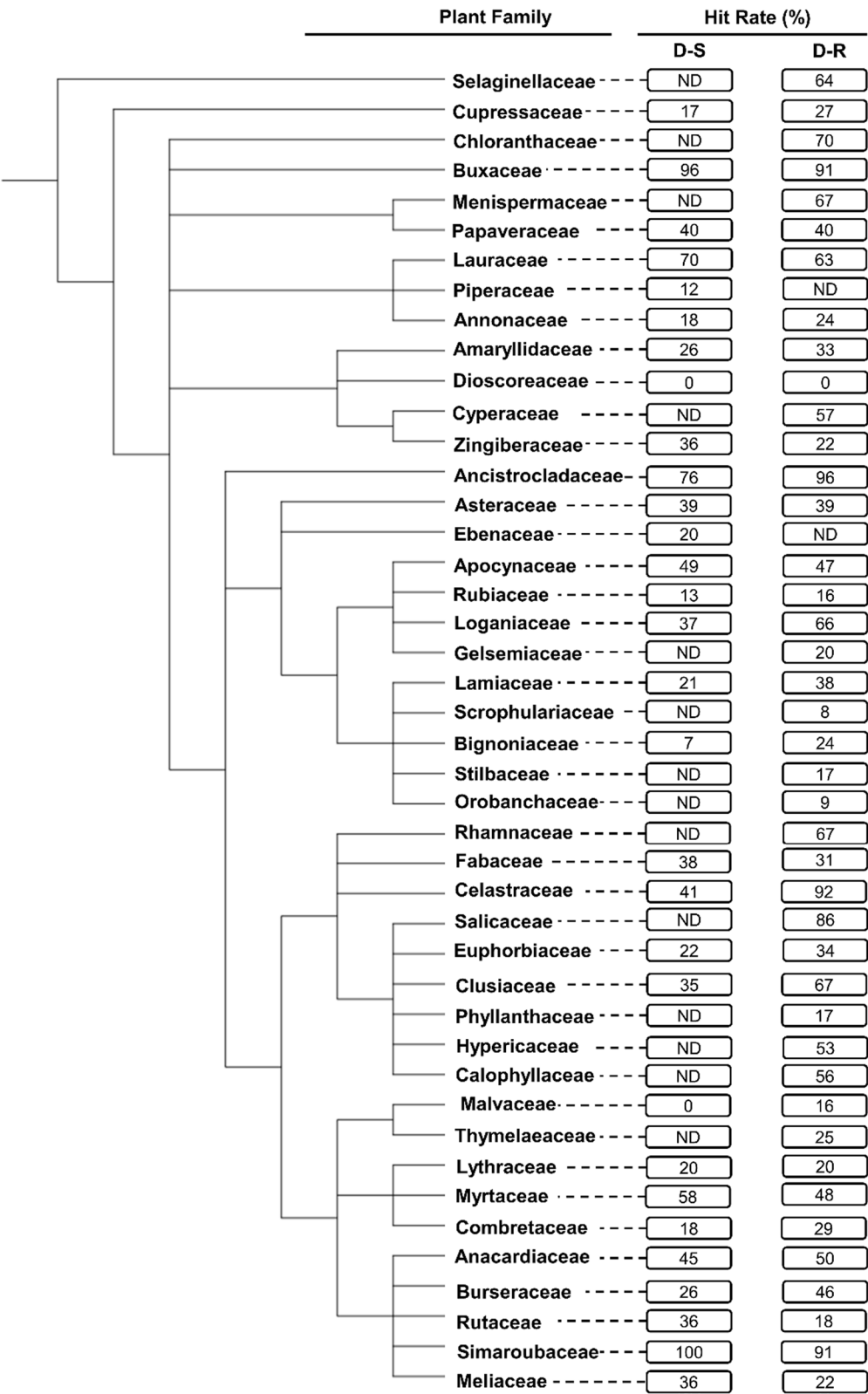


Fig. 3 Phylogenetic tree of plant families investigated in vitro for their activity against the intra-erythrocytic asexual *P. falciparum* parasites. The tree was generated using NCBI Taxonomy and processed using iTOL. ND not determined: This applies to plant families with < 10 compounds isolated from them and subsequently evaluated for their antiplasmodial activity. The hit rate (HR) is the % of compounds with an $IC_{50} \leq 10 \mu M$ for each plant family

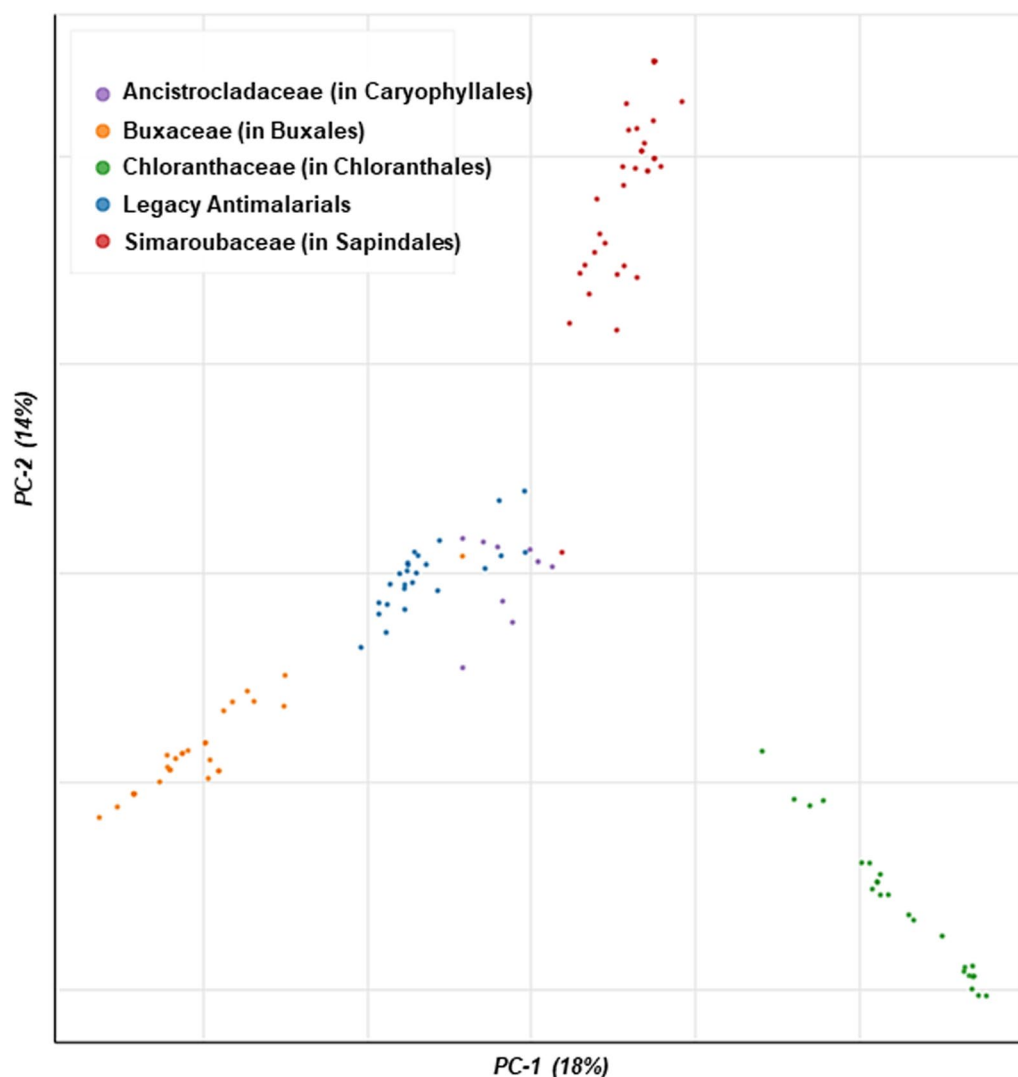


Fig. 4 Launched drug chemical space of the 'legacy' antimalarials and natural product compounds isolated from the 'hot' plant orders and families. The online Python library for chemical space visualization ChemPlot [33], was used to launch the chemical space of the natural compounds and 'legacy' antimalarials

D-S *P. falciparum* strains. The productivity of the Ancistrocladaceae ($n=45$) and Simaroubaceae ($n=55$) was retained against D-R strains with 49 and 62% of compounds from each respective plant family being classified as HA. Further plant families with most compounds (ranging from 45 to 75%) classified as HA against D-R were the Buxaceae ($n=11$), Chloranthaceae (in Chloranthales) ($n=44$), Celastraceae (in Celastrales) ($n=12$), and Loganiaceae (in Gentianales) (41). Intriguingly, most compounds (63%) from the Loganiaceae ($n=84$), were classified as PA against the D-S parasites. Despite receiving considerable research interest (as observed by the number of compounds isolated from them and screened for their antiparasmodial activity), many compounds

isolated from the families Fabaceae (in Fabales), Rutaceae (in Sapindales), Rubiaceae (in Gentianales), Annonaceae (in Magnoliales) and Asteraceae (in Asterales) were classified as PA against both D-S and D-R *P. falciparum* parasites (Table 4).

The resistance index for all plant families is good since >90% of compounds have an RI < 10. Complementing the good activity profile of most compounds in the Ancistrocladaceae ($n=43$), Buxaceae ($n=31$), and Chloranthaceae ($n=17$) was a good SI for most of them (>60%) (Table 4).

From this expanded analysis, plants of the Buxales and Caryophyllales along with those in the families Simaroubaceae, Buxaceae, and Ancistrocladaceae have

Table 3 Antiplasmodial activity and cytotoxicity of compounds isolated from plants of different orders

Plant Order		Inhibition of parasite proliferation								Cytotoxicity			
		D-S				D-R							
		n	HA	MA	PA	n	HA	MA	PA			n	RI (≤10)
Asterids	Lycopodiales					11	18	45	36			7	57
	Cupressales	25	4	16	80	16	0	31	69	6	100	5	60
	Ranunculales	21	33	29	38	45	20	42	38	12	91.6	33	27
	Chloranthales					44	55	16	30			13	85
	Buxales	25	36	60	4	11	46	46	9			31	65
	Magnoliales	91	3	13	84	117	7	15	78	32	100	46	22
	Laurales	12	8	58	33	21	24	38	38			14	43
	Piperales	32	0	25	75	15	33	13	53	8	100	18	61
	Dioscoreales	14	0	0	100	14	0	0	100				
	Asparagales	31	6	26	68	35	23	23	54	16	100	35	26
	Zingiberales	11	9	27	64	18	0	22	78	7	85.7	6	17
	Poales					16	0	50	50				
	Caryophyllales	53	55	25	21	62	45	37	18	39	97.4	49	92
	Cornales	11	27	0	73							6	33
	Ericales	23	0	13	87							13	8
	Apiales	10	0	40	60							8	3
	Asterales	151	5	34	61	145	2	37	61	47	98	146	16
	Gentianales	192	11	19	70	190	17	23	60	94	98.9	108	35
	Lamiales	56	5	20	75	98	4	20	76	11	100	64	28
Rosids	Brassicales	10	20	20	80	10	10	0	90			8	50
	Sapindales	168	14	25	61	287	16	24	60	77	94.8	158	
	Malvales	13	15	8	77	31	3	13	81	7	100	10	40
	Myrtales	53	11	26	62	88	3	31	66	27	100	46	37
	Fagales	19	21	11	68	21	0	38	62	10	90	13	31
	Malpighiales	86	8	28	64	186	9	41	49	32	93.7	113	15
	Rosales					28	0	61	39	5	100	13	8
	Oxalidales	13	0	31	69	10	0	20	80	10	90	9	0
	Fabales	138	8	30	62	177	9	21	69	66	100	86	42
Celastrales	22	36	5	59	12	75	17	8	10	100	11	45	
		0	50	100	HM								

0 50 100 HM

n = number of compounds. HA, MA and PA values are expressed in % of total compounds evaluated. RI shows % of compounds with RI ≤ 10 . SI shows % of compounds with SI ≥ 10 . For RI and SI we have only presented data where ≥ 5 compounds have been evaluated and reported. The heat map (HM) ranges from green (lowest value, 0%) to yellow (mid-range value, 50%) to red (highest value, 100%), visually illustrating the proportion of compounds classified as either HA, MA or PA

retained their 'hot' status. In addition to these, plants of the orders Chloranthales and Celastrales and the families Chloranthaceae and Celastraceae emerge as 'hot' taxonomic plant groups, particularly in producing potent compounds against the D-R *Plasmodium* parasites. It is quite striking to note that despite being most represented, the asterids and rosids (two major eudicot groups) only have one 'hot' plant order (the Celastrales) and two 'hot' families (the Simaroubaceae and Celastraceae) emerging from these lineages. It is noted though that generally, the HR for most compounds isolated from taxa in these clades is still relatively high, and noteworthy in relation to antimalaria drug discovery.

2.4 Drug-likeness assessment of compounds produced by different higher plant orders and families

Potent antiplasmodial compounds should have good drug-like properties for ease of development into orally available preclinical and clinical candidates with reduced attrition rates in clinical trials. Drug-like properties include physicochemical descriptors including for example molecular weight (MW), consensus LogP (cLogP), hydrogen bond donors (HBD) and acceptors (HBA). Preference for drug discovery should be given to plant orders and families that produce compounds with good drug-like properties. To assess the drug-likeness of compounds isolated from the different plant orders and families, in silico calculated molecular and

Table 4 Activity and cytotoxicity of compounds isolated from plant species in respective plant families

Plant Family		Inhibition of parasite proliferation										Cytotoxicity				
		D-S				D-R						n	RI (≤10)	n	SI (≥10)	
		n	HA	MA	PA	n	HA	MA	PA							
Ranu	Selaginellaceae					11	18	46	36			7	57.1			
	Cupressaceae	24	4	13	83	15	0	27	73	5	100					
	Chloranthaceae					44	55	16	30			13	84.6			
	Buxaceae	25	36	60	4	11	46	46	9			31	64.5			
	Menispermaceae					33	21	45	33			30	26.6			
	Papaveraceae	10	0	40	60	10	0	40	60							
	Lauraceae	10	10	60	30	19	26	37	37			12	41.6			
	Piperaceae	26	0	12	88					8	100	9	22.2			
	Annonaceae	84	4	14	82	102	8	16	76	25	100	41	24.3			
	Amaryllidaceae	19	0	26	74	24	8	25	67	12	100	21	23.8			
	Dioscoreaceae	14	0	0	100	14	0	0	100							
	Cyperaceae					14	0	57	43							
	Zingiberaceae	11	9	27	64	18	0	22	78	7	85.7	6	16.6			
	Ancistrocladaceae	42	52	24	24	45	49	47	4	30	100	43	93			
	Gent	Asteraceae	151	5	34	61	145	2	37	61	47	98	146	16		
Ebenaceae		15	0	20	80											
Apocynaceae		37	24	24	51	77	16	31	53	18	100	46	43.4			
Rubiaceae		71	4	8	87	61	5	11	84	38	100	38	26.3			
Loganiaceae		84	11	26	63	41	41	24	34	38	97.3	18	50			
Gelsemiaceae						10	0	20	80			5	20			
Lamiales		Lamiaceae	28	7	14	79	45	4	33	62			44	22.7		
		Scrophulariaceae					25	0	8	92			9	44.4		
		Bignoniaceae	14	0	7	93	17	0	24	76	6	83.3	13	30.7		
		Stilbaceae					12	0	17	83						
		Orobanchaceae					11	0	9	91						
		Rhamnaceae					24	0	67	33	5	100	13	7.6		
		Fabaceae	138	8	30	62	177	9	21	69	66	100	86	41.8		
		Celastraceae	22	36	5	59	12	75	17	8	10	100	11	45.5		
		Malpighiales	Salicaceae					14	21	64	14	7	100	13	0	
	Euphorbiaceae		18	11	11	78	29	17	17	66	6	100	11	36.3		
	Clusiaceae		48	2	33	65	72	13	54	33	16	93.7	51	17.6		
	Phyllanthaceae						18	0	17	83			12	16.7		
	Hypericaceae						17	0	53	47			14	7.1		
	Calophyllaceae						16	0	56	44						
	Malv		Malvaceae	10	0	0	100	19	0	16	84	5	100	7	14.2	
Thymelaeaceae							12	8	17	75						
Lythraceae			10	0	20	80	10	0	20	80						
Myrt			Myrtaceae	26	19	38	42	27	7	41	52	12	100	16	37.5	
			Combretaceae	17	6	12	82	51	2	27	71	11	100	28	39.2	
			Sapindales	Anacardiaceae	11	9	36	55	10	20	30	50	8	100	7	0
				Burseraceae	23	0	26	74	26	0	46	54	5	80	41	31.7
				Rutaceae	77	8	29	64	98	5	13	82	31	100	39	35.8
				Simaroubaceae	19	74	26	0	55	62	29	9	13	84.6	35	62.8
		Meliaceae		37	5	14	81	93	4	28	68	23	100	35	25.7	
					0	50	100	HM								

HM

n = number of compounds. HA, MA and PA values are expressed in % of total compounds evaluated. RI shows % of compounds with RI ≤ 10 . SI shows % of compounds with SI ≥ 10 . For RI and SI we have only presented data where ≥ 5 compounds have been evaluated and reported. *Myrt* Myrtales, *Malv* Myrtales, *Gent* Gentianales, *Ranu* Ranunculales. The heat map (HM) ranges from green (lowest value, 0%) to yellow (mid-range value, 50%) to red (highest value, 100%), visually illustrating the proportion of compounds classified as either HA, MA or PA

physicochemical descriptors of compounds were evaluated using different sets of criteria and by utilising data of clinically available antimalarial drugs (Tables 6 and 7). The analysis showed that a significant portion of these compound descriptors agreed with those of the criteria outlined by the Medicines for Malaria Venture, Lipinski's Rule of 5 [27], Veber's rules [28] and Ghose filters [29] indicating good characteristics of drug-likeness. However, some compounds isolated from the Buxales, Chloranthales, and Caryophyllales did not wholly fulfil the set criteria (Table 5). These discrepancies were noted for the respective families, which

included the Buxaceae, Chloranthaceae and Ancistrocladaceae (Table 6). Out of the seven obtained physicochemical descriptors for compounds in these plant orders and families, some molecules did not fall within the specified set criteria for MW, the HBA, molar refractivity (MR), and cLogP.

Further evaluation showed average descriptor values of many of the compounds from most plant families and orders compared well with those of approved antimalarial drugs. Similarly, not all antimalarial drug descriptors fell within the criteria for drug-likeness. For example, lumefantrine has a MW of 528.9, a MR of 152.6, cLogP

Table 5 Calculated mean physicochemical descriptors for compounds isolated from different plant orders*

Plant Order	Physicochemical descriptors							PAINS	SA
	MW	RB	HBA	HBD	MR	TPSA	cLogP		
Lycopodiales	547.4	3.9	10.0	5.1	147.5	169.3	3.8	0.0	4.5
Cupressales	317.3	2.4	2.8	1.6	94.1	50.2	4.1	0.1	4.3
Ranunculales	426.2	2.0	5.7	0.5	124.0	56.2	3.7	0.1	4.6
Chloranthales	599.9	5.9	10.2	2.5	151.2	154.1	2.5	0.0	7.3
Buxales	503.0	5.3	4.0	2.0	149.7	60.2	5.6	0.0	6.4
Magnoliales	334.5	4.9	4.6	1.5	93.9	68.8	3.1	0.1	3.9
Laurales	345.7	4.1	4.7	0.9	98.8	55.0	3.1	0.0	4.2
Piperales	353.2	6.2	4.2	2.2	102.8	72.6	3.9	0.3	3.6
Dioscoreales	340.8	2.6	6.8	4.2	88.2	121.8	1.4	0.6	4.1
Asparagales	409.3	3.6	6.9	2.2	109.4	97.1	2.1	0.2	4.7
Zingiberales	307.6	3.9	3.5	1.2	87.2	55.2	3.2	0.0	5.1
Poales	271.3	2.1	3.6	1.8	77.2	57.7	2.9	0.0	3.7
Caryophyllales	497.5	4.2	7.0	3.2	147.2	95.8	4.2	0.3	5.0
<i>Asterids</i>									
Cornales	433.6	3.7	3.5	1.5	130.2	59.8	5.2	0.0	5.5
Ericales	517.1	4.3	6.6	3.6	143.2	109.4	4.3	0.1	5.7
Apiales	524.7	6.7	7.1	4.2	143.3	115.8	3.7	0.0	6.7
Asterales	392.9	5.6	6.2	2.1	104.9	96.5	2.8	0.1	5.1
Gentianales	393.0	2.9	4.4	1.7	115.5	67.6	2.8	0.3	4.8
Lamiales	374.0	4.6	6.3	3.1	98.8	101.9	2.0	0.3	4.8
<i>Rosids</i>									
Brassicales	430.0	6.4	7.1	3.6	118.6	112.4	2.7	0.1	4.9
Sapindales	410.2	4.4	6.1	1.8	111.2	91.4	2.9	0.1	5.1
Malvales	417.2	4.2	7.6	3.4	110.1	117.2	2.6	0.1	4.1
Myrtales	448.8	3.1	7.3	3.9	120.7	122.7	2.9	0.3	5.2
Fagales	548.9	7.8	10.2	4.9	140.7	166.1	2.3	0.4	6.1
Malpighiales	461.3	5.4	6.4	2.8	130.3	106.5	4.3	0.3	5.1
Rosales	520.8	6.1	6.0	2.6	148.1	102.6	4.8	0.1	5.7
Oxalidales	519.8	11.4	12.2	7.1	127.1	205.8	1.2	0.5	5.1
Fabales	361.1	4.1	5.3	1.8	101.0	77.9	3.2	0.1	4.2
Celastrales	512.9	8.3	4.6	2.6	151.3	79.2	5.9	0.0	6.4

MW molecular weight, RB rotatable bonds, HBA hydrogen bond acceptors, HBD hydrogen bond donors, MR molar refractivity, TPSA total polar surface area, cLogP consensus LogP, PAINS pan-assay interference compounds, SA synthesis accessibility

*Italic figures are those which don't meet the set criteria

Table 6 Calculated mean physicochemical descriptors for compounds isolated from different plant families*

Plant Family	Physicochemical descriptors							PAINS	SA
	MW	RB	HBA	HBD	MR	TPSA	cLogP		
Selaginellaceae	547.4	3.9	10.0	5.1	147.5	169.3	3.8	0.0	4.5
Cupressaceae	317.3	2.4	2.8	1.6	94.1	50.2	4.1	0.1	4.3
Chloranthaceae	599.9	5.9	10.2	2.5	151.2	154.1	2.5	0.0	7.3
Buxaceae	503.0	5.3	4.0	2.0	149.7	60.2	5.6	0.0	6.4
<i>Ranu</i>									
Menispermaceae	461.0	2.2	6.1	0.6	136.0	58.1	4.0	0.0	5.1
Papaveraceae	343.5	1.2	5.4	0.5	95.7	55.5	2.8	0.1	3.3
Lauraceae	347.7	4.2	4.8	0.9	99.2	55.6	3.1	0.0	4.3
Piperaceae	354.2	7.1	4.0	2.3	104.5	70.8	4.3	0.3	3.6
Annonaceae	336.8	4.8	4.7	1.5	94.1	71.3	3.0	0.1	3.9
Amaryllidaceae	311.9	1.4	5.2	1.1	85.2	61.5	1.5	0.0	4.3
Dioscoreaceae	340.8	2.6	6.8	4.2	88.2	121.8	1.4	0.6	4.1
Cyperaceae	287.9	2.0	3.9	1.9	81.1	63.1	3.0	0.0	3.8
Zingiberaceae	307.6	3.9	3.5	1.2	87.2	55.2	3.2	0.0	5.1
Ancistrocladaceae	507.4	4.1	6.3	2.5	154.9	79.2	5.0	0.2	5.0
Asteraceae	392.9	5.6	6.2	2.1	104.9	96.5	2.8	0.1	5.1
Ebenaceae	378.6	3.2	2.7	1.3	113.4	47.1	5.2	0.1	4.8
<i>Gent</i>									
Apocynaceae	421.3	3.7	4.1	1.4	125.9	62.5	3.4	0.2	5.0
Rubiaceae	339.9	3.3	5.6	2.1	91.8	86.8	2.3	0.3	4.0
Loganiaceae	418.5	1.7	3.3	1.4	130.3	49.2	3.0	0.3	5.3
Gelsemiaceae	358.4	3.2	5.8	2.2	96.9	92.6	1.4	0.0	5.2
<i>Lamiales</i>									
Lamiaceae	302.6	2.7	4.2	1.8	84.0	68.0	2.6	0.5	4.2
Scrophulariaceae	494.0	8.0	10.6	5.9	119.9	169.2	0.1	0.1	6.2
Bignoniaceae	509.0	6.8	9.5	5.0	129.8	156.9	1.8	0.5	6.0
Stilbaceae	434.0	5.3	3.9	1.8	128.1	69.4	5.4	0.3	5.4
Orobanchaceae	365.1	4.3	7.7	4.5	90.5	127.1	0.7	0.2	4.8
Rhamnaceae	550.1	6.4	6.4	2.6	155.8	107.3	5.0	0.1	6.0
Fabaceae	361.1	4.1	5.3	1.8	101.0	77.9	3.2	0.1	4.2
Celastraceae	512.9	8.3	4.6	2.6	151.3	79.2	5.9	0.0	6.4
<i>Malpighiales</i>									
Salicaceae	520.8	11.6	7.5	0.8	142.3	100.8	4.7	0.0	6.9
Euphorbiaceae	471.2	5.6	8.8	2.9	122.5	140.4	2.4	0.3	5.1
Clusiaceae	465.8	5.4	6.3	3.4	134.3	110.4	4.8	0.4	5.0
Phyllanthaceae	511.0	7.7	6.5	3.3	143.7	112.6	4.3	0.1	5.7
Hypericaceae	383.7	3.9	5.2	2.7	110.9	88.3	3.9	0.3	3.7
Calophyllaceae	419.5	1.9	4.6	2.1	123.4	77.9	5.0	0.1	4.6
<i>Malv</i>									
Malvaceae	368.7	4.1	6.5	2.4	99.0	95.1	2.7	0.1	3.8
Thymelaeaceae	472.1	4.3	9.2	4.8	121.1	149.6	2.2	0.1	4.5
<i>Myrt</i>									
Lythraceae	529.5	1.6	11.4	5.5	137.7	182.5	2.0	0.4	5.6
Myrtaceae	455.1	3.3	6.8	2.8	123.7	112.8	3.5	0.3	5.7
Combretaceae	430.5	3.1	6.8	4.3	115.7	118.6	2.6	0.2	4.9
<i>Sapindales</i>									
Anacardiaceae	389.4	8.7	4.8	3.0	112.0	85.6	4.1	0.2	4.9
Burseraceae	337.7	2.0	2.8	1.2	101.1	45.5	4.4	0.1	5.0
Rutaceae	340.5	4.0	5.0	1.2	94.9	71.1	2.8	0.1	3.7
Simaroubaceae	459.6	2.9	9.3	3.6	110.9	147.1	0.4	0.0	6.2
Meliaceae	512.8	6.6	7.2	1.5	138.8	106.6	3.9	0.0	6.5

Table 6 (continued)

MW molecular weight, *RB* rotatable bonds, *HBA* hydrogen bond acceptors, *HBD* hydrogen bond donors, *MR* molar refractivity, *TPSA* total polar surface area, *cLogP* consensus LogP, *Myrt* Myrtales, *Malv* Myrtales, *Gent* Gentianales, *Ranu* Ranunculales, *PAINS* pan-assay interference compounds, *SA* synthesis accessibility

*Italics figures are those which do not meet the set criteria. *Myrt* Myrtales, *Malv* Myrtales, *Gent* Gentianales, *Ranu* Ranunculales

Table 7 Ranking of plant orders for antimalarial drug discovery

Plant Order			Points					Ranking		
			D-S		D-R		SI		Total	Normalised
			HR	HA	HR	HA				
Asterids	Lycopodiales			5	9	6	20	6.7	5	
	Cupressales	22	19	18	20	5	84	16.8	21	
	Ranunculales	4	4	6	8	17	39	7.8	7	
	Chloranthales			4	2	2	8	2.7	3	
	Buxales	1	3	2	3	3	12	2.4	2	
	Magnoliales	23	20	21	15	19	98	19.6	26	
	Laurales	3	13	7	6	9	38	7.6	6	
	Piperales	19	21	11	5	4	60	12.0	10	
	Dioscoreales	25	21	26	20		92	23.0	29	
	Asparagales	14	16	12	7	18	67	13.4	13	
	Zingiberales	12	12	21	20	20	85	17.0	22	
	Poales			10	20		30	15.0	18	
	Caryophyllales	2	1	3	4	1	11	2.2	1	
	Cornales	18	5			14	37	12.3	11	
	Ericales	24	21	26	20	23	114	22.8	28	
	Apiales	6	21			25	52	17.3	23	
	Asterales	8	18	15	19	21	81	16.2	19	
	Gentianales	17	11	14	10	13	65	13.0	12	
	Lamiales	19	17	20	16	16	88	17.6	25	
Rosids	Brassicales	6	7	25	12	7	57	11.4	9	
	Sapindales	9	9	13	11		42	10.5	8	
	Malvales	21	8	24	18	11	82	16.4	20	
	Myrtales	11	10	17	17	12	67	13.4	13	
	Fagales	15	6	16	20	15	72	14.4	17	
	Malpighiales	13	14	9	13	22	71	14.2	16	
	Rosales			8	20	24	52	17.3	23	
	Oxalidales	16	21	23	20	26	106	21.2	27	
	Fabales	10	15	19	14	10	68	13.6	15	
Celastrales	5	2	1	1	8	17	3.4	4		
			1	29	RSHM					

HR Hit rate. Ranking score heat map (RSHM) ranges from red 'hot' (Best ranking, lowest points) to green (Lowest ranking, most points)

of 7.9 and doxycycline and tetracycline both have 6 HBD and a TPSA of 181.6 Å².

Encouragingly, compounds from most of the plant orders and families were shown to be devoid of the

pan-assay interference compounds (PAINS) sub-structures. The synthesis accessibility (SA) of many compounds from most plant orders and families was consistent with much of the currently available antimalarials.

Notable exceptions were the compounds in the orders Buxales, Chloranthales, and Caryophyllales and the families Buxaceae, Chloranthaceae and Ancistrocladaceae. Their SA (ranging from 5 to 7.3) was in the same range as those of the artemisinin derivatives (6.5 to 6.7) (Tables 5 and 6). Most appealing will be plant orders and families which produce compounds which with a low SA value are easy to synthesise, exemplified by the quinolines.

2.5 Overall ranking to identify 'hot' higher plant orders and families for prioritisation in drug discovery projects

We formulated a rational ranking system which we used to identify the 'hottest' plant orders and families. We awarded points to different plant orders and families depending on how they performed in three attributes: the hit rate, HA for both D-S and D-R parasites, and the SI. We decided against including RI and drug-likeness in the ranking system as most of the compounds from most plant orders and families largely complied with set criteria for these indicators, so markedly reducing the value of these characteristics in improving selection resolution. Points were sequentially awarded based on the position of the taxonomic group in performance relative to other groups. For example, 1 point was given, per performance indicator, to the order or family with the highest hit rate, HA, and SI. The second, and third-positioned taxonomic groups were given 2 and 3 points, respectively. This scoring was repeated in that sequence until points were assigned to all plant families and orders. Thereafter, the total number of points was calculated for each family and order. We finally rationalised the number of points based on the number of indicators scored per taxonomic group resulting in the normalised points. This was done to 'level' the system, as not all taxonomic groups were scored for all indicators. We then ranked the taxonomic groups based on the number of points with the family or plant order, with the lowest number of normalised points ranking 1st and that with the greatest number of points ranking last (Tables 7 and 8 for orders and families respectively).

Having adopted this ranking system, the following results were obtained and found to be consistent with earlier observations: the top 3 ranked orders were the (i) Caryophyllales, (ii) Buxales, and (iii) Chloranthales. The top 3 ranked plant families were the (i) Ancistrocladaceae (in Ancistrocladaceae), (ii) Simaroubaceae (in Sapindales), and (iii) Buxaceae (in Buxaceae) (Tables 7 and 8). The most prominent natural product classes found to be active ($IC_{50} \leq 10 \mu M$) per each plant order and family were isoquinoline alkaloids and naphthoquinones (Caryophyllales and Ancistrocladaceae), steroid alkaloids and lupane triterpenoids (Buxales and Buxaceae), quassinoids

(Simaroubaceae) and cycloodesmane sesquiterpenoids (Chloranthales) (Table 9). We point out that majority of compounds classified by NPClassifier are manually classified as naphthylisoquinoline (NIQs) alkaloids in their respective research publications, most of which emanate from the research group of Professor G. Bringmann [30]. Encouragingly, the HA compounds from these top plant taxa are structurally different to the 'legacy' antimalarial drugs [31].

The pressing need to discover and develop new antimalarials to mitigate drug resistance led us to consider the use of phylogenetic analysis coupled with bioactivity correlation to establish 'hot' plant orders and families worthy of prioritisation for antimalarial drug discovery projects. This endeavour has culminated in the identification of 3 'hot' plant orders and families. One of the most intriguing findings of the current study was that the most promising plant orders and families are those from which either no antimalarial drug has previously been isolated or they are generally less studied (judging by either number of compounds isolated from them or number of publications reporting their investigation). In contrast, the families Asteraceae and Rubiaceae have received significant interest in their antiplasmodial evaluation. This interest we believe is driven by two factors, (i) previous discovery of antimalaria drugs from these families and (ii) their extensive use traditionally for malaria treatment as documented by ethnobotanical studies (e.g., [34, 35], and [36]). However, with a combined total of *ca.* 400 compounds isolated from the Asteraceae and Rubiaceae plant families, it is striking to note that only 16 of them (*ca.* 4%) have demonstrated IC_{50} values $\leq 1 \mu M$ either against D-S or D-R *Plasmodium* parasites. In contrast, of the less investigated plant families Simaroubaceae, Ancistrocladaceae, and Buxaceae, with a combined total of only 86 compounds isolated and screened for activity against the D-S *Plasmodium* strains, 45 of these compounds (53%) were reported to show IC_{50} values of $\leq 1 \mu M$. Furthermore, the compounds isolated from these 3 families and those from other 'hot' taxonomic groups show good selectivity, outperforming the Asteraceae and Rubiaceae families (see Table 4).

From a medicinal chemistry perspective, emphasis is placed on the discovery of new compounds that occupy a chemical space that is different from that of the current clinically available antimalarial drugs. This chemical diversity brings with it a high likelihood of targeting novel biological space [37]. Having a unique target, compared to current antimalarials, increases the chances of the new scaffolds being potent against clinically drug-resistant *Plasmodium* strains. Our analysis shows that the HA compounds isolated from the 'hot' plant orders and families identified from the study occupy a different

Table 8 Ranking of plant families for antimalarial drug discovery

		Points						Ranking	
		D-S		D-R		SI	Total		Normalised
		HR	HA	HR	HA				
Ranu	Selaginellaceae			11	11	5	27	9.0	8
	Cupressaceae	25	19	28	26		98	24.5	35
	Chloranthaceae			6	3	2	11	3.7	4
	Buxaceae	2	4	3	5	3	17	3.4	3
	Menispermaceae			7	9	18	34	11.3	10
	Papaveraceae	9	22	20	26		77	19.3	18
	Lauraceae	4	9	12	7	11	43	8.6	7
	Piperaceae	27	22			24	73	24.3	32
	Annonaceae	23	20	30	18	21	112	22.4	30
	Amaryllidaceae	16	22	24	16	22	100	20.0	20
	Dioscoreaceae	29	22	42	26		119	29.8	42
	Cyperaceae			13	26		39	19.5	19
	Zingiberaceae	13	10	32	26	28	109	21.8	29
	Ancistrocladaceae	3	2	1	4	1	11	2.2	1
	Asteraceae	10	17	21	24	29	101	20.2	24
	Ebenaceae	20	22				42	21.0	26
Gent	Apocynaceae	6	5	18	13	9	51	10.2	9
	Rubiaceae	26	18	38	21	19	122	24.4	34
	Loganiaceae	12	8	10	6	6	42	8.4	6
	Gelsemiaceae			33	26	25	84	28.0	39
Lamiales	Lamiaceae	19	14	22	22	23	100	20.0	20
	Scrophulariaceae			41	26	8	75	25.0	37
	Bignoniaceae	28	22	30	26	17	123	24.6	36
	Stilbaceae			37	26		63	31.5	43
	Orobanchaceae			40	26		66	33.0	44
	Rhamnaceae			7	26	31	64	21.3	28
Malpighiales	Fabaceae	11	12	26	15	10	74	14.8	12
	Celastraceae	8	3	2	1	7	21	4.2	5
	Salicaceae			5	8	33	46	15.3	15
	Euphorbiaceae	18	7	23	12	14	74	14.8	12
	Clusiaceae	15	21	7	14	26	83	16.6	16
	Phyllanthaceae			36	26	27	89	29.7	41
	Hypericaceae			15	26	32	73	24.3	32
	Calophyllaceae			14	26		40	20.0	20
	Malvaceae	29	22	39	26	30	146	29.2	40
	Thymelaeaceae			29	17		46	23.0	31
Myrt	Lythraceae	20	22	33	26	33	134	26.8	38
	Myrtaceae	5	6	17	19	13	60	12.0	11
	Combretaceae	24	15	27	25	12	103	20.6	25
Sapindales	Anacardiaceae	7	10	16	10	33	76	15.2	14
	Burseraceae	17	22	19	26	16	100	20.0	20
	Rutaceae	13	13	35	20	15	96	19.2	17
	Simaroubaceae	1	1	4	2	4	12	2.4	2
	Meliaceae	22	16	25	23	20	106	21.2	27
		1	44	RSHM					

Table 8 (continued)

HR Hit rate. Ranking score heat map (RSHM) ranges from red 'hot' (Best ranking, lowest points) to green (Lowest ranking, most points). *Myrt* Myrtales, *Malv* Myrtales, *Gent* Gentianales, *Ran* Ranunculales

Table 9 Natural compound classification of active compounds in 'hot' plant orders and families[#]

Natural product class (NPC)	Plant Order (% NPC composition)			Plant Family (% NPC composition)		
	Caryophyllales	Buxales	Chloranthales	Ancistrocladaceae	Simaroubaceae	Buxaceae
Unclassified*	17.1			21.9		
Unclassified [#]		4.2				4.2
Flavonols	2.4					
Isoquinoline alkaloids	43.9			40.6		
Naphthalenes and derivatives	2.4			37.5		
Naphthoquinones	31.7					
Tetraketide meroterpenoids	2.4					
Steroidal alkaloids		75.0				75.0
Lupane triterpenoids		12.5				12.5
Pregnane steroids		8.3				8.3
Cycloeudesmane ^ψ			100.0			
Quassinoids					100.0	

*# Compounds could only be assigned to Pathway and Super Class using the online tool NPCClassifier [32] and not the natural product class. ^ψCycloeudesmane – cycloeudesmane sesquiterpenoids

chemical space to current antimalarials providing further impetus to explore these taxonomic groups. For example, the most prolific compounds from the Simaroubaceae are classed as quassinoids, which the parasite has clinically not been exposed to [38]. Similarly, from the Buxaceae family, the most prolific compounds are steroidal alkaloids, which are chemically distinct from any of the clinically available curative malaria drugs [39].

The quassinoids class of natural products, which the Simaroubaceae is well-established to produce, has also demonstrated in vivo antiplasmodial activity albeit with some level of toxicity noted. For example, the quassinoid bruceine B was shown to have an ED₉₀ of 2.82 mg/kg/day. At a concentration threefold the ED₉₀, bruceine B was observed to be 100% lethal against the mice used in the study [38]. Nonetheless, this compound was shown to be less toxic than other quassinoids, so raising hope that through medicinal chemistry less toxic but highly potent scaffolds from this natural product class could be synthesised. Notably, the synthesis of potent yet less in vivo toxic quassinoid analogues has been successfully undertaken for cancer studies [40]. NIQs have similarly shown exceptional in vivo activity. The NIQ dioncophylline C cured *P. bhergei*-infected mice following a single oral dose (50 mg/kg/day) with no observed toxicity [41]. While NIQs are structurally highly complex, approaches to their synthesis have been developed and comprehensively outlined [42] by the group of Professor G. Bringmann [30]. Moreover, simplified analogues of this class

of compounds have been synthesised and proved to be potent against intra-erythrocytic asexual *Plasmodium* parasites [43]. Their clinical efficacy remains to be demonstrated.

It is interesting to note the generally high HR of most natural products isolated from many plant orders and families discussed in this study. These HR were substantially higher than those observed for synthetic compounds which have been described to be as low as 0.3% and 0.05% in some studies [44, 45]. However, caution needs to be exercised when considering these high hit rates for natural products. Firstly, the bioassay-guided assay approach is a popular approach used to isolate compounds from plants, where guidance is based on the observed bioactivity resulting in the isolation of bioactive molecules, albeit with varying potency. Secondly, the cut-off point (IC₅₀ ≤ 10 μM) for the HR outlined in this paper is noted to be more tolerant than that used elsewhere, e.g., < 1.25 μM and ≤ 2 μM adopted by Plouffe et al. [45] and Decherling et al. [44], respectively. Nevertheless, the high HR is still encouraging, motivating for the continued investigation of plant-derived compounds to treat malaria.

In conclusion, given the need to accelerate antimalarial drug discovery, plants are a promising oasis deserving of continued investigation in this endeavor. Our study has shown that understudied plant orders and families are more deserving of intensified investigation in search of novel antimalarial drugs. We anticipate these findings

will help direct researchers to focus and streamline their investigations on the few plant orders and families most likely to result in the discovery of highly active antiparasmodial compounds that can be channeled into medicinal chemistry programs.

3 Material and methods

3.1 Literature search

To identify published manuscripts for exploration in our study, we queried the PubMed database (<https://pubmed.ncbi.nlm.nih.gov/>) [46] searching for publications documenting antiparasmodial properties of compounds isolated from plants. The key phrase used was “*Plasmodium falciparum* and natural product” limiting the “Text Availability” option to ‘Abstract’. The search was restricted to manuscripts published between 1964 and 2021. We then manually systematically screened the publications applying the following exclusion criteria.

- i) Manuscripts documenting only antiparasmodial activity of compounds isolated from other natural sources, e.g., microorganisms, marine organisms, other than vascular (i.e., higher) plants were discarded.
- ii) Manuscripts in which no compounds were isolated and screened were disregarded.
- iii) Manuscripts in which only in vivo studies were carried out were excluded.
- iv) Duplicate articles were excluded.

Articles subsequently remaining following the above process, were selected for the study.

3.2 Taxonomic terminology

From the selected manuscripts, we manually collated compounds reported along with relevant pharmacological data, and species-of-origin saving this information on a Microsoft Excel spreadsheet. The captured data was verified 3 times to ensure all details collected were accurate and consistent with current nomenclature and taxonomy. Given disparities in taxon circumscriptions and related nomenclature inherent at all taxon ranks, we harmonised our systematic approach through aligning with the World Flora Online (WFO) database (<http://www.worldfloraonline.org/>) as of 7 November 2022, including in the assignment of species authors. In instances where the species authors for taxa had not been provided in the source pharmacological publications it was occasionally necessary, when more than one identical basionym exists, to resolve the identity of the research subject through consideration of the reported plant collection locality relative to data provided in the Global Biodiversity Information Facility (GBIF) (<https://www.gbif.org/>).

3.3 Phylogenetic tree generation and data analysis

The phylogenetic trees were constructed as follows. Firstly, Text (.txt) files with the names of plant orders and families (as per WFO) were added onto the NCBI Taxonomic Browser ‘Common Tree’ [24]. The resulting tree was saved as a ‘Phylogeny file’ (.phy) which was graphically displayed and manipulated using the iTOL online tool (v5) [25]. Here, default settings were used with only the following modifications made; Branch lengths—‘Ignore’ and Scaling factors—‘0.5 × horiz.’

The SMILES of the compounds collated were either collected from databases including PubChem [47] and ChemSpider [48] or were generated from 2D structures drawn on ChemDraw Ultra (v8) [49]. Using the generated SMILES, compounds were classified into specific classes using the NPClassifier tool [32]. To evaluate drug-likeness we computed average physicochemical properties using the SwissADME online suite software [50]. Analysis, including calculation of mean values for RI, SI etc., was carried out using Microsoft Excel. Chemical space analysis was carried out on ChemPlot using structural similarity, PCA algorithm and scatter plot type options [33].

Acknowledgements

This research work was supported by a grant from the Department of Science and Innovation (DSI) of South Africa awarded to Vinesh Maharaj. The UP ISMC acknowledges the South African Medical Research Council as Collaborating Centre for Malaria Research. Phanankosi Moyo was supported by a grant from the University of Pretoria Postgraduate Research Support Bursary for his Postdoctoral Fellowship. Sephora Mianda Mutombo was supported by funds from the University of Pretoria Postgraduate Research Support Bursary, South Africa and the L’Oréal-UNESCO for Woman in Science grant. Luke Invernizzi was supported by funds from the doctoral funds from National Research Foundation of South Africa.

Author Contribution

Conceptualisation of study, PM and VJM; Collecting data sets, PM, LI, SMM, WR, and AWA; Formal analysis, PM, AWA, WR, MW and NRC; Writing—original draft, PM; Writing—review and editing, PM, VJM, LI, SMM, AWA, MW, and NRC. Supervision and funds acquisition VJM; All authors read and approved the final manuscript.

Funding

Foundation L’Oréal (4500453975).

Availability of data and materials

The data that support the findings of this study are available from the corresponding author upon reasonable request.

Declarations

Competing interests

MW is a co-founder of Ometa Labs LLC.

Received: 3 August 2023 Accepted: 3 September 2023
Published online: 05 October 2023

References

- Cox FE. History of the discovery of the malaria parasites and their vectors. *Parasit Vectors*. 2010;3(1):1–9.
- WHO. World malaria report 2022. 2022: World Health Organization.
- Ippolito MM, et al. Antimalarial drug resistance and implications for the WHO global technical strategy. *Curr Epidemiol Rep*. 2021;8:46–62.
- Matthews H, Duffy CW, Merrick CJ. Checks and balances? DNA replication and the cell cycle in *Plasmodium*. *Parasit Vectors*. 2018;11(1):216.
- Gupta DK, et al. DNA damage regulation and its role in drug-related phenotypes in the malaria parasites. *Sci Rep*. 2016;6(1):23603.
- Blasco B, Leroy D, Fidock DA. Antimalarial drug resistance: linking *Plasmodium falciparum* parasite biology to the clinic. *Nat Med*. 2017;23(8):917–28.
- Newman DJ, Cragg GM. Natural products as sources of new drugs over the nearly four decades from 01/1981 to 09/2019. *J Nat Prod*. 2020;83(3):770–803.
- Wells TNC. Natural products as starting points for future anti-malarial therapies: going back to our roots? *Malar J*. 2011;10(1):53.
- Yang J, et al. Advances in the research on the targets of anti-malaria actions of artemisinin. *Pharmacol Ther*. 2020;216: 107697.
- Institute of Medicine Committee on the Economics of Antimalarial D. In Arrow KJ, Panosian C, Gelband H, editors. Saving lives, buying time: economics of malaria drugs in an age of resistance. National Academies Press (US); 2004. Copyright 2004 by the National Academy of Sciences. All rights reserved.: Washington (DC).
- Milliken W, et al. Plants used traditionally as antimalarials in Latin America: mining the tree of life for potential new medicines. *J Ethnopharmacol*. 2021;279: 114221.
- Christenhusz MJ, Byng JW. The number of known plants species in the world and its annual increase. *Phytotaxa*. 2016;261(3):201–17.
- Douwes E, et al. Regression analyses of southern African ethnomedicinal plants: informing the targeted selection of bioprospecting and pharmacological screening subjects. *J Ethnopharmacol*. 2008;119(3):356–64.
- Holzmeyer L, et al. Evaluation of plant sources for anti-infective lead compound discovery by correlating phylogenetic, spatial, and bioactivity data. *Proc Natl Acad Sci*. 2020;117(22):12444–51.
- Zhu F, et al. Clustered patterns of species origins of nature-derived drugs and clues for future bioprospecting. *Proc Natl Acad Sci*. 2011;108(31):12943–8.
- Rønsted N, et al. Phylogenetic selection of *Narcissus* species for drug discovery. *Biochem Syst Ecol*. 2008;36(5–6):417–22.
- Mawalagedera SM, et al. Combining evolutionary inference and metabolomics to identify plants with medicinal potential. *Front Ecol Evol*. 2019;7:267.
- Prasad MA, Zolnik CP, Molina J. Leveraging phytochemicals: the plant phylogeny predicts sources of novel antibacterial compounds. *Fut Sci OA*. 2019;5(7):FSO407.
- Mahajan GB, Balachandran L. Antibacterial agents from actinomycetes—a review. *Front Biosci-Elite*. 2012;4(1):240–53.
- Aminov R. History of antimicrobial drug discovery: major classes and health impact. *Biochem Pharmacol*. 2017;133:4–19.
- Berkov S, et al. Chemodiversity, chemotaxonomy and chemoecology of Amaryllidaceae alkaloids. *Alkaloids Chem Biol*. 2020;83:113–85.
- Li X, et al. What makes species productive of anti-cancer drugs? Clues from drugs' species origin, druglikeness, target and pathway. *Anti-Cancer Agents Med Chem (Formerly Current Medicinal Chemistry-Anti-Cancer Agents)*. 2019;19(2):194–203.
- Egieyeh SA, et al. Prioritization of anti-malarial hits from nature: chemoinformatic profiling of natural products with in vitro antiplasmodial activities and currently registered anti-malarial drugs. *Malar J*. 2016;15:1–23.
- Schoch CL, et al. NCBI Taxonomy: a comprehensive update on curation, resources and tools. *Database*. 2020. 2020.
- Letunic I, Bork P. Interactive Tree Of Life (iTOL) v5: an online tool for phylogenetic tree display and annotation. *Nucleic Acids Res*. 2021;49(W1):W293–6.
- Borsch T, et al. World Flora Online: placing taxonomists at the heart of a definitive and comprehensive global resource on the world's plants. *Taxon*. 2020;69(6):1311–41.
- Lipinski CA, et al. Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Adv Drug Deliv Rev*. 2001;46:3–26.
- Veber DF, et al. Molecular properties that influence the oral bioavailability of drug candidates. *J Med Chem*. 2002;45(12):2615–23.
- Ghose AK, Viswanadhan VN, Wendoloski JJ. A knowledge-based approach in designing combinatorial or medicinal chemistry libraries for drug discovery. 1. A qualitative and quantitative characterization of known drug databases. *J Comb Chem*. 1999;1(1):55–68.
- Trauner D, Roßmann K. A conversation with Gerhard Bringmann. *ACS Publications*; 2020. p. 1248–1252.
- Charman SA, et al. An in vitro toolbox to accelerate anti-malarial drug discovery and development. *Malar J*. 2020;19(1):1–27.
- Kim HW, et al. NPClassifier: a deep neural network-based structural classification tool for natural products. *J Nat Prod*. 2021;84(11):2795–807.
- Cihan Sorkun M, et al. ChemPlot, a Python library for chemical space visualization. *Chemistry-Methods*. 2022;2(7): e202200005.
- Lianaa D, Rungsihirunrata K. Using phylogeny approach on ethnobotanical bioprospecting for leading antimalarial plant-based drug discovery.
- Nondo RS, et al. Ethnobotanical survey and in vitro antiplasmodial activity of medicinal plants used to treat malaria in Kagera and Lindi regions, Tanzania. *J Med Plants Res*. 2015;9(6):179–92.
- Yetein MH, et al. Ethnobotanical study of medicinal plants used for the treatment of malaria in plateau of Allada, Benin (West Africa). *J Ethnopharmacol*. 2013;146(1):154–63.
- Hovlid ML, Winzeler EA. Phenotypic screens in antimalarial drug discovery. *Trends Parasitol*. 2016;32(9):697–707.
- O'Neill MJ, et al. Plants as sources of antimalarial drugs, part 4: activity of *Brucea javanica* fruits against chloroquine-resistant *Plasmodium falciparum* in vitro and against *Plasmodium berghei* in vivo. *J Nat Prod*. 1987;50(1):41–8.
- Szabó LU, et al. Antiprotozoal nor-triterpene alkaloids from *Buxus sempervirens* L. *Antibiotics*. 2021;10(6):696.
- Pei Y, et al. Quassinoid analogs with enhanced efficacy for treatment of hematologic malignancies target the PI3K γ isoform. *Commun Biol*. 2020;3(1):267.
- Francois G, et al. Naphthylisoquinoline alkaloids against malaria: evaluation of the curative potentials of dioncophylline C and dioncopeltine A against *Plasmodium berghei* in vivo. *Antimicrob Agents Chemother*. 1997;41(11):2533–9.
- Tajuddeen N, et al. The stereoselective total synthesis of axially chiral naphthylisoquinoline alkaloids. *Acc Chem Res*. 2022;55(17):2370–83.
- Bringmann G, et al. Synthesis and antiprotozoal activities of simplified analogs of naphthylisoquinoline alkaloids. *Eur J Med Chem*. 2008;43(1):32–42.
- Decherer KJ, et al. Replenishing the malaria drug discovery pipeline: screening and hit evaluation of the MMV Hit Generation Library 1 (HGL1) against asexual blood stage *Plasmodium falciparum*, using a nano luciferase reporter read-out. *SLAS Discov*. 2022;27(6):337–48.
- Plouffe D, et al. In silico activity profiling reveals the mechanism of action of antimalarials discovered in a high-throughput screen. *Proc Natl Acad Sci*. 2008;105(26):9059–64.
- White J. PubMed 2.0. *Medical Reference Services Quarterly*. 2020. 39(4): 382–387.
- Kim S, et al. PubChem substance and compound databases. *Nucleic Acids Res*. 2016;44(D1):D1202–13.
- Pence HE, Williams A. ChemSpider: an online chemical information resource. *ACS Publications*; 2010.
- Mendelsohn LD. ChemDraw 8 ultra, windows and macintosh versions. *J Chem Inf Comput Sci*. 2004;44(6):2225–6.
- Daina A, Michielin O, Zoete V. SwissADME: a free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules. *Sci Rep*. 2017;7(1):42717.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.